

## New Medical Strategies for Midgut Carcinoids

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**Abstract:** Patients with well-differentiated neuroendocrine tumours of the gastrointestinal tract often present with metastases and hormonal symptoms. These patients can be palliated by interventional tumour reduction and medical treatment with somatostatin analogues; no effective chemotherapy is available. Radionuclide therapy via somatostatin receptors is one new therapeutic alternative. The recognition that neuroendocrine tumours express specific receptors for growth factors and chemokines, which are of importance for tumour growth, vascularization, and spread, may open the way for new therapeutic approaches. The signalling pathways in carcinoid tumours are incompletely explored. This review summarizes potential new treatment strategies from clinical and experimental studies, e.g. inhibition of angiogenesis, targeting of growth factors or their receptors by tyrosine kinase inhibitors, interference with specific cellular pathways (mTOR, PI3K, RAS/RAF, Notch), and also inhibition of the proteasome and histone deacetylation. Combining targeted therapy with chemotherapy, or using drugs to sensitize for radionuclide therapy, may enhance the treatment outcome.

**Keywords:** Angiogenesis inhibition, growth factor receptor, monoclonal antibody, multikinase inhibition, neuroendocrine tumour, radionuclide therapy, small molecule inhibitor, somatostatin receptor.

### INTRODUCTION

Gastrointestinal neuroendocrine tumours can be divided into endocrine pancreatic tumours (EPTs) and carcinoids of the tubular gastrointestinal tract, of which ileal (midgut) carcinoids are the most common. Both tumour types are usually well-differentiated, with the capacity to synthesize and release biogenic amines and peptide hormones that cause specific symptoms, which can be controlled to a certain degree by long-acting somatostatin analogues.

Surgical resection is the only curative treatment of carcinoid tumours; patients with distant metastases are less often accessible for such procedures and may be subject to tumour reduction accomplished by debulking surgery, hepatic arterial embolization, or radio-frequency ablation [1]. In carefully selected patients, liver transplantation may be considered [2]. For widely disseminated disease, radionuclide therapy via highly expressed somatostatin receptors (SSTR) on the tumours, or combinations of chemotherapeutic agents or biotherapy with interferons have been attempted, but with limited success. Patients with residual metastatic disease can suffer from severe hormonal symptoms. It is therefore of some urgency to find new medical strategies to reduce tumour growth and alleviate the hormonal symptoms. Clinical studies on molecularly targeted and anti-angiogenic therapies have been started and the early results offer some promise. There have been few experimental studies on signalling pathways in midgut carcinoid tumours, and they still lack confirmation in the clinical situation.

### CLINICAL THERAPEUTIC PRINCIPLES

#### Targeting of Somatostatin Receptors

Somatostatin receptors were first detected in the CNS and the pituitary, and were later found to be localized in the gastrointestinal tract with high expression in a large proportion of neuroendocrine tumours [3]. Somatostatin reduces smooth muscle contractility and glandular secretion by inhibiting cAMP. Reduced tumour growth was observed early in studies of cell lines [4, 5], which was attributed to hyperphosphorylation of the RB gene product and cell cycle arrest. Five subtypes of SSTR have been cloned, which differ in tissue distribution but also between tumour types, i.e. midgut carcinoids have high expression of SSTR2 and SSTR5 [6]. With the

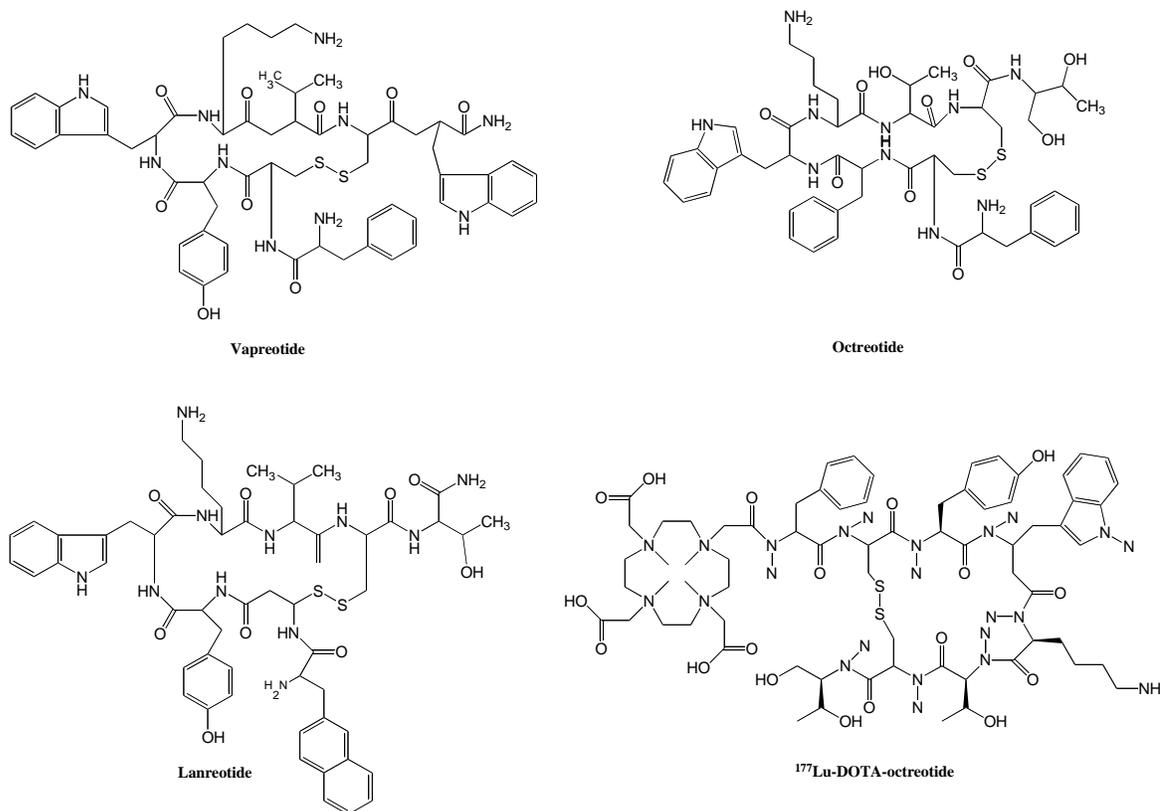
synthesis of cyclic somatostatin analogues (*octreotide*, *vapreotide*, and *lanreotide*), which are resistant to peptidases, these compounds were shown to have much longer half-lives and biological activity than native somatostatin (Fig. 1). The analogues had high affinity to SSTR2 and SSTR5, medium affinity to SSTR3, and low affinity to SSTR1 and SSTR4 [7]. A pan-analogue (SOM 230) was developed with high affinity to all SSTRs except SSTR4 [8]. The formation of hetero-oligomers targeting dopamine- and somatostatin receptors with enhanced functional activity was also shown early on [9].

The anti-secretory effect of somatostatin analogues has been most useful to reduce carcinoid symptoms (facial flush and diarrhoea) caused by hyper-secretion of serotonin and tachykinins, and to prevent carcinoid crisis during interventional treatment [10]. Recently, an anti-proliferative effect by the depot formulation octreotide LAR was first shown in randomized studies of patients with limited tumour burden of midgut carcinoids [11]. Radiation therapy, mediated through  $\beta$ -emitting radionuclides bound to somatostatin analogues that are internalized into tumour cells after receptor binding, is one attractive way of treating disseminated disease [12] and may be an adjunct to intentionally curative surgery in the case of micrometastases (Fig. 1). Tailored chemotherapy for several tumour types based on their expression of peptide receptors is currently under investigation [13].

#### Targeting of EGFR and PDGFR

Therapy targeting specific growth factors and their receptors, e.g. inhibition of epithelial growth factor receptor (EGFR) or vascular endothelial growth factor (VEGF) pathways in advanced colon cancer, or of the EGFR tyrosine kinase in non-small cell lung carcinoma (NSCLC) after failure with chemotherapy, has encouraged the use of similar principles for neuroendocrine tumours. These tumours can express both receptors for platelet-derived growth factor (PDGF) and VEGF and their ligands as part of autocrine/paracrine loops [14, 15]. In immunohistochemical studies, VEGF was found to be present in half of the EPTs studied; and EGFR, the tyrosine kinase receptor KIT (with stem cell factor as ligand), and basic fibroblast growth factor (bFGF) were present in about one quarter. The pattern differed from that of carcinoids, which almost invariably expressed EGFR, and VEGF also in more than half of the tumours [16]. EGFR expression was frequently upregulated and it was suggested to influence the growth and spread of these tumours [17]. PDGFA and PDGFB may be expressed in both tumour and stromal tissue [14].

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**Fig. (1).** Three commonly used somatostatin analogues. Radiolabelled octreotate is used for radionuclide therapy.

The first clinical studies targeting tyrosine kinases were performed in almost 100 patients with progressive disease after treatment using the EGFR tyrosine kinase inhibitor (TKI) *gefitinib* [18] (Fig. 2). Radiological tumour regression was observed in less than 10% of cases, but tumour stabilization was more frequent; it was notably more common in patients with carcinoids than in patients with EPT (the 6-month progression-free survival was 30% vs. 10%). In an ongoing trial of advanced gastrointestinal cancer, including carcinoids, another EGFR-TKI, *erlotinib*, was combined with the EGFR-targeted antibody cetuximab (to tag tumour cells with IgG for the immune effector cells) (NIH: NCT 00397384). In studies of other tumour types, EGFR targeting appeared to have optimal effects first when combined with other targeted therapies or chemotherapy [19-21]. Hobday and collaborators<sup>1</sup> studied a multi-targeted TKI, *sorafenib* (VEGFR2-3, PDGFR, KIT, the Fms-Like Tyrosine kinase 3 (FLT3), the oncoprotein BRAF, and FGFR1) leading to radiological RECIST (Response Evaluation Criteria in Solid Tumours) responses in about 10% of the patients, which were equally frequent in carcinoids and EPT. Due to the complex actions of this drug, it can be difficult to ascribe the observed effects to interference with the oncogenic pathway RAF/MEK/ERK, anti-angiogenic activity, or other mechanisms. It must be borne in mind that when signalling pathways are inhibited in tumour cells, alternative pathways can be activated, leading to reduced therapeutic effects [22]. The spontaneous growth of neuroendocrine tumours may vary considerably, which means that disease stabilization can be difficult to evaluate if data on tumour progression prior to therapy are not available. Another problem with phase-II studies is that

recruited patients can have considerable selection bias due to numerous previous treatments.

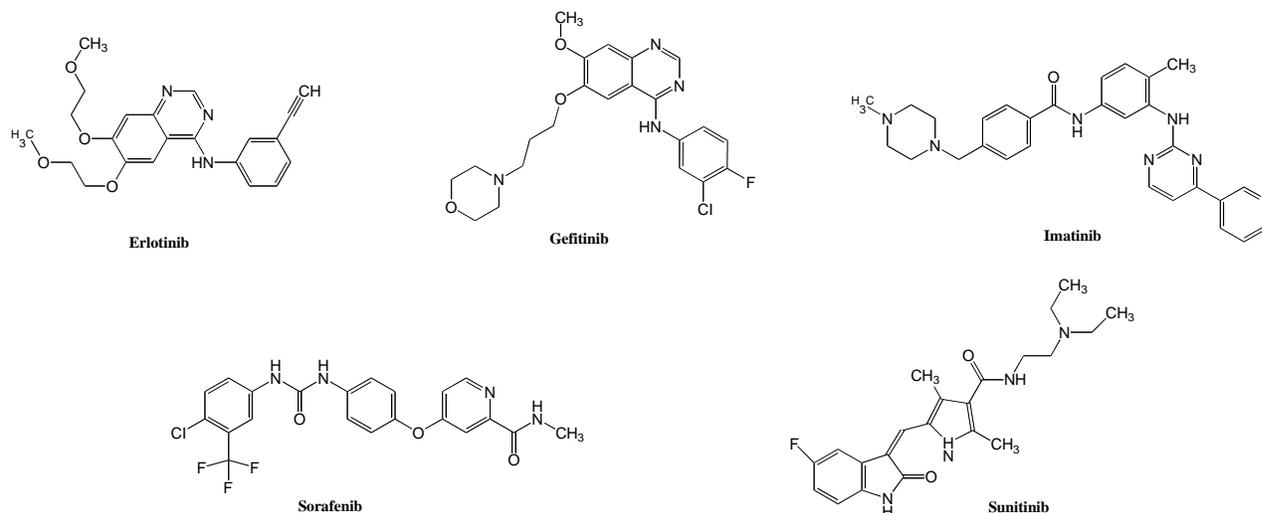
*Sunitinib* is a multi-target TKI (VEGFR1-3, PDGFR, KIT, the oncoprotein RET, and FLT3), which was also studied in almost 100 patients with progressive neuroendocrine tumour disease (Fig. 2). Also here, tumour regression by RECIST criteria was unusual (16% for EPT but only 2% for carcinoids), while tumour stabilization was much more frequent (75–93%) [23]. A phase-III trial in patients with advanced EPT was recently stopped due to differences in efficacy; the sunitinib-treated group had a progression-free survival of 11.1 months as opposed to 5.5 months in the placebo group<sup>2</sup>. Preliminary results on sunitinib combined with hepatic arterial embolization are also encouraging<sup>3</sup>.

*Imatinib*, the first TKI tailored for treatment of chronic myeloid leukemia and gastrointestinal stromal tumours (GIST), also targets PDGFR, which may be expressed by carcinoids. This drug has weak effects on PDGFR in comparison with sunitinib and has no effect on VEGFR (Fig. 2). Accordingly, in limited studies of advanced carcinoids only one patient showed objective responses; drug-related toxicity was seen due to high dosage [24, 25]. It can still not be excluded that imatinib has a role as an adjunct to VEGF inhibition [26]. For EPT and carcinoids, specific activating mutations of target kinases cannot be related to therapeutic outcome, i.e. a success similar to that of imatinib in GIST should not be expected. With knowledge of the mutational status of critical target

<sup>1</sup>Hobday, T. J.; Rubin, J.; Holen, K.; Picus, J.; Donehower, R.; Marschke, R.; Maples, W.; Lloyd, R.; Mahoney, M.; Erllichman, C. MC044h, a phase II trial of sorafenib in patients (pts) with metastatic neuroendocrine tumors (NET): A Phase II Consortium (P2C) study. *J. Clin. Oncol. (Meeting Abstracts)*, **2007**, *25*(18 suppl), 4504.

<sup>2</sup>Raymond, E.; Raoul, J.; Niccoli, P.; Bang, Y. J.; Borbath, I.; Lombard-Bohas, C.; metrakos, P.; Lu, D. R.; Blankmeister, C.; Vinik, A. Phase III, randomized, double-blind trial of sunitinib vs placebo in patients with progressive well-differentiated malignant pancreatic islet cell tumours. *Ann. Oncol.*, **2009**, *20*(Suppl 7), vii11.

<sup>3</sup>Strosberg, J. R.; Campos, T.; Kvoils, L. K. Phase II study of sunitinib maleate following hepatic artery embolization for metastatic gastroenteropancreatic endocrine tumors. *ASCO (Meeting Abstracts)*, **2009**; p. 272.



**Fig. (2).** Small-molecule inhibitors of EGFR (gefitinib, erlotinib) and multi-target TKIs (sorafenib, sunitinib). Imatinib was the first TKI to target KIT and PDGFRA.

genes in GIST, the choice of an optimal drug can be made, e.g. imatinib for patients with *KIT* exon 11 mutations, sunitinib for *KIT* exon 9 mutations, or in the case of drug resistance due to acquired tyrosine kinase mutations a multi-target TKI can be used [27]. Similar principles can be used to some extent for other tumour types, e.g. NSCLC. The demonstration of mutational activation of *EGFR* in these tumours is helpful in selecting patients who will respond to gefitinib, while a high number of *EGFR* gene copies characterizes patients who are responsive to erlotinib [28].

### Targeting of mTOR

The protein kinase mammalian target of rapamycin (mTOR) is a downstream regulator of signalling from several growth factors, e.g. via the phosphatidylinositol 3 kinase (PI3K)/AKT pathway, and influences the early protein translation that is a prerequisite for cell cycle progression. Under normal conditions, PI3K/AKT is controlled by a suppressor gene, phosphatase and tensin homologue (*PTEN*). When *PTEN* is inactivated by mutations, epigenetic events, or altered subcellular localization of its protein, mTOR is activated and can participate in the tumorigenesis [29, 30]. It is therefore of interest that phosphorylation of AKT has been observed in neuroendocrine tumours and that inhibition of mTOR induces anti-proliferative effects *in vitro* [29, 31]. Modern mTOR inhibitors (*everolimus* and *temsirolimus*) are analogues of the natural antibiotic rapamycin (*sirolimus*) that was originally used for immunosuppressive treatment in patients undergoing organ transplants (Fig. 3). They have been used with some success as rescue therapy in patients with GIST and acquired TKI-resistance [27]. In a limited study, *everolimus* was combined with octreotide LAR (to reduce autocrine/paracrine secretion of growth factors) in patients with EPT or carcinoids. Chromogranin A (CgA) is a granular neurosecretory protein released together with tumour-specific hormones; it serves as a general plasma marker of neuroendocrine tumour disease. After treatment with everolimus, the CgA levels were reduced and long, progression-free intervals were seen; the 6-month progression-free survival was 64%. Again, EPT appeared to be more responsive than carcinoids (27% as opposed to 17%) and the treatment was well tolerated [32]. The confirmatory phase-II trials in progressive refractory EPT have recently been reported (RADIANT 1) [33]. The RADIANT 2 study is directed at advanced carcinoids.

### Inhibition of Angiogenesis

Antiangiogenic therapy has recently been attempted in phase-II studies of patients with neuroendocrine tumours, since these tu-

mours are often well vascularized and frequently express VEGF (or its subtypes VEGF B-E) and/or the placental growth factors (PlGF1 and PlGF2) [15, 34]. Furthermore, there is a correlation between circulating levels of VEGF and progression of these tumours [35] and also between elevated expression of VEGF and both enhanced angiogenesis and reduced progression-free survival [36]. Also, other angiogenic factors, e.g. bFGF and PDGF sub-types, can be released from tumour cells or tumour vasculature. When VEGF binds to high-affinity receptors on the endothelium (VEGFR1 and VEGFR2), or on lymph vessels (VEGFR3), the intracellular domain of the receptor becomes phosphorylated. This in turn activates downstream signalling, leading to angiogenesis. Inhibition of this process can be accomplished with antibodies to VEGF or its receptors, or with TKI, which inhibits receptor phosphorylation. The second generation of TKIs after imatinib (*sunitinib* and *sorafenib*) target several tyrosine kinases, including those of VEGFR1–VEGFR3, and they therefore inhibit angiogenesis also. Recombinant endostatin mimics the endogenous anti-angiogenesis, while metalloproteinase inhibitors influence the degradation of the extracellular matrix, which is an early step in angiogenesis.

*Bevacizumab* (a monoclonal antibody to VEGF) has a proven therapeutic effect when combined with chemotherapy in colon cancer [37]. In a phase-II trial, bevacizumab was tested as monotherapy vs. pegylated interferon- $\alpha$  for advanced carcinoids; bevacizumab led to improved progression-free survival and markedly reduced tumour perfusion<sup>4</sup>. When octreotide was combined with bevacizumab, patients with carcinoids more frequently demonstrated early tumour remissions than after treatment with pegylated interferon- $\alpha$  and octreotide [38]. When bevacizumab was combined with the alkylating agent *temozolomide* (a dacarbazine analogue), partial remissions and stable disease were seen more often in patients with EPT than in patients with carcinoids, which may reflect differences in vascularization between the tumour types<sup>5</sup>. When bevacizumab was combined with standard treatment of gastrointestinal cancer in phase-II trials of patients with neuroendocrine tu-

<sup>4</sup> Yao, J. C.; Ng, C.; Hoff, P. M.; Phan, A. T.; Hess, K.; Chen, H.; Wang, X.; Abbruzzese, J. L.; Ajani, J. A. Improved progression free survival (PFS), and rapid, sustained decrease in tumor perfusion among patients with advanced carcinoid treated with bevacizumab. *J. Clin. Oncol. (Meeting Abstracts)*, **2005**, 23(16 suppl), 4007.

<sup>5</sup> Kulke, M. H.; Stuart, K.; Earle, C. C.; Bhargava, P.; Clark, J. W.; Enzinger, P. C.; Meyerhardt, J.; Attawia, M.; Lawrence, C.; Fuchs, C. S. A phase II study of temozolomide and bevacizumab in patients with advanced neuroendocrine tumors. *J. Clin. Oncol. (Meeting Abstracts)*, **2006**, 24(18 suppl), 4044.

mours, e.g. capecitabine and oxaliplatin or FOLFOX (5-fluorouracil, leucovorin, and oxaliplatin), 20–30% of the patients showed partial responses<sup>6,7</sup>. Bevacizumab has also been investigated in patients with advanced carcinoids in combination with 2-methoxyestradiol (NIH: NCT00328497). The latter substance can reduce growth and induce apoptosis in both endothelium and tumour cells, and inhibit hypoxia-inducible factor (HIF-1), which drives pro-angiogenic genes [39, 40].

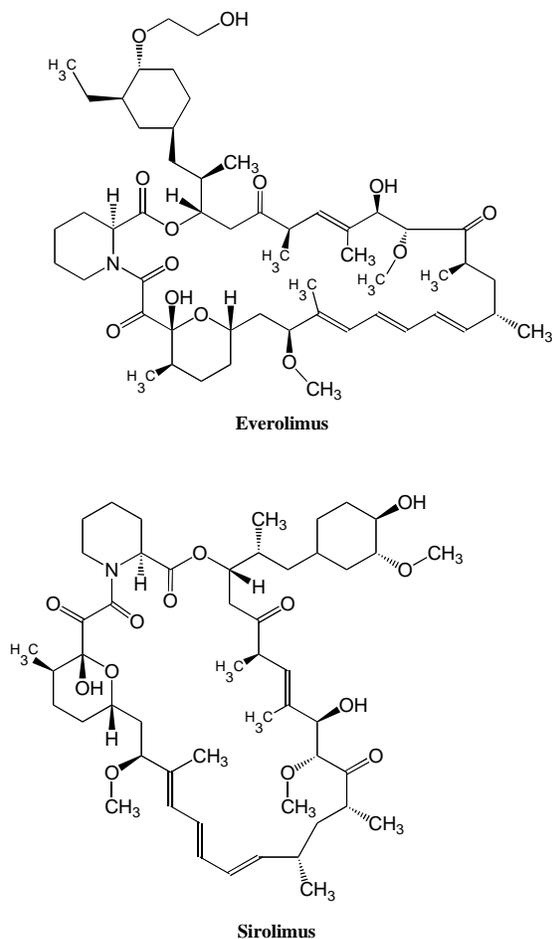


Fig. (3). Inhibitors of mTOR.

*Vatalanib* inhibits all VEGFR tyrosine kinases and might therefore block both tumour angiogenesis and lymphangiogenesis (Fig. 4). Relatively few patients with neuroendocrine tumours have been studied in phase-I and phase-II studies, demonstrating no radiological responses so far, but with a high frequency of disease stabilization<sup>8,9</sup>. Common side effects (nausea and vomiting) were mild, but hypertension occurred in individual patients.

<sup>6</sup> Kunz, P. L.; Kuo, T.; Kaiser, H. L.; Norton, J. A.; Longacre, T. A.; Ford, J. M.; Fisher, G. A. A phase II study of capecitabine, oxaliplatin, and bevacizumab for metastatic or unresectable neuroendocrine tumors: Preliminary results. *J. Clin. Oncol. (Meeting Abstracts)*, **2008**, 26(15 suppl), 15502.

<sup>7</sup> Venook, A. P.; Ko, A. H.; Tempero, M. A.; Uy, J.; Weber, T.; Korn, M.; Bergsland, E. K. Phase II trial of FOLFOX plus bevacizumab in advanced, progressive neuroendocrine tumors. *J. Clin. Oncol. (Meeting Abstracts)*, **2008**, 26(15 suppl), 15545.

<sup>8</sup> Anthony, L.; Chester, M.; Michael, S.; O'Dorisio, T. M.; O'Dorisio, M. S. Phase II open-label clinical trial of vatalanib (PTK/ZK) in patients with progressive neuroendocrine cancer. *J. Clin. Oncol. (Meeting Abstracts)*, **2008**, 26(15 suppl), 14624.

<sup>9</sup> Pavel, M. E.; Bartel, C.; Heuck, F.; Neumann, F.; Tiling, N.; Pape, U. F.; Plockinger, U.; Wiedenmann, B. Open-label, non-randomized, multicenter phase II study evaluating the angiogenesis inhibitor PTK787/ZK222584 (PTK/ZK) in patients with

Two pan-VEGFR inhibitors, *pazopanib* and *AMG706*, have been developed with additional inhibition of PDGFR and KIT [41, 42] (Fig. 4). Monotherapy with *AMG706* is presently tested in patients with low-grade neuroendocrine tumours to assess tolerability and early anti-tumour effects (NIH:NCT00427349), while *pazopanib* has reached phase-II studies (NIH:NCT00454363).

In animal studies, anti-PIGF monoclonal antibodies were shown to block the angiogenic switch in tumours without influencing the physiological vasculature and may be useful in cases of resistance to VEGF inhibitors [43]. The antibodies also appear to block rescue-angiogenesis, which can be a problem in long-term anti-angiogenic therapy.

*Thalidomide* interferes not only with VEGF and bFGF signaling, but also with the formation of extracellular matrix. *Thalidomide* is therefore a multifaceted anti-angiogenic drug. Together with *temozolomide*, it has been used in a phase-II trial in 29 patients with neuroendocrine tumours. The overall radiological response rate was 25% and tumour stabilization was almost three times higher, but at the cost of high toxicity (fatigue, thrombocytopenia, neuropathy) [44] (Fig. 4).

Recombinant endostatin (a 20-kD fragment of collagen XVIII) was tested in a phase-II trial of 42 patients with neuroendocrine tumours. No radiological responses were observed, but 80% showed stable disease for a median of 11 months and the toxicity was low [45].

Anti-angiogenic therapy is thus a new approach of some promise in neuroendocrine oncology. In the studies cited, partial tumour responses were unusual in patients with carcinoids (< 10%), but appeared to be about twice as common in patients with EPT.

## EXPERIMENTAL THERAPEUTIC PRINCIPLES

The study of neuroendocrine tumours has been hampered by a number of circumstances, which were recently reviewed [46]. The main obstacles to progress in gastroenteropancreatic neuroendocrine tumour management include (1) our limited understanding of the cellular and molecular biology of neuroendocrine cells and the mechanisms of tumorigenesis, (2) a shortage of *in vitro* and animal models to study disease pathogenesis and treatment, (3) a paucity of critical targets for new therapies, and (4) a lack of uniform pathological classification and staging systems. The lack of a uniform classification system is evident in most clinical studies on neuroendocrine tumours. The carcinoid tumour group is heterogeneous and comprises a mixture of foregut-, midgut-, and hindgut-derived tumours. The clinical response rate of carcinoid tumours is often compared to that of EPTs, which are also a heterogeneous group of tumours with marked differences in clinical presentation and malignant behaviour. Furthermore, clinical studies are commonly performed on patients with tumours displaying varying degrees of differentiation, proliferation rates, and disease stage, thus limiting the validity of the conclusions drawn. Experimental data from well-characterized *in vitro* and animal models are therefore needed to properly evaluate novel treatment principles for ileal carcinoids. To date, five human carcinoid cell lines have been characterized: the pancreatic carcinoid cell line BON [47] and the ileal (midgut) carcinoid cell lines KRJ-1 [48], GOT1 [49], CNDT2 [50], and STS [51]. Three of the cell lines (BON, KRJ-1, and GOT1) have been used to identify therapeutic targets; only two models, KRJ-1 and GOT1, are representative of enterochromaffin cell (ileal) carcinoids [52, 53]. In the following sections, we give different examples of anti-tumour effects demonstrated in carcinoid cells *in vitro* and in animal models; these effects suggest possible new treatment principles for ileal carcinoids.

advanced neuroendocrine carcinomas (NEC). *J. Clin. Oncol. (Meeting Abstracts)*, **2008**, 26(15 suppl), 14684.

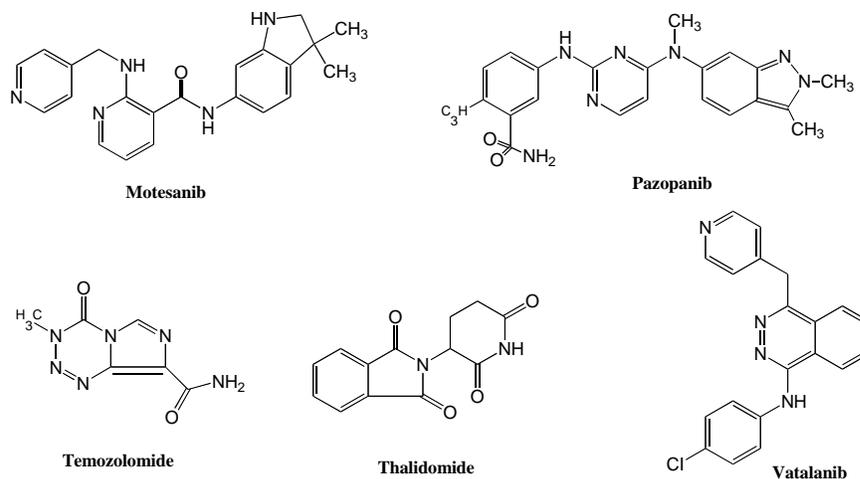


Fig. (4). Inhibitors of angiogenesis; motesanib, pazopanib, and vatalanib are TKIs.

### Targeting of PARP

Nuclear factor- $\kappa$ B (NF- $\kappa$ B)/Rel transcription factors are important regulators of both inflammation and tumorigenesis. The tumorigenic effects of activated NF- $\kappa$ B include promotion of malignant transformation, proliferation, inhibition of apoptosis, invasiveness, angiogenesis, and metastasis formation [54]. Several human tumour types have been shown to harbour constitutively activated NF- $\kappa$ B [55-60] and in some cases mutations associated with NF- $\kappa$ B signalling have been found [61]. The most effective way of interfering with NF- $\kappa$ B activation is to inhibit the activity of the I $\kappa$ B (inhibitory protein of NF- $\kappa$ B) kinase (IKK) [62] (Fig. 5). CHS828 (or GMX1778) is a pyridyl guanidine with anti-tumour activity *in vitro* and *in vivo*. Earlier studies have shown that CHS828 inhibits the activation of NF- $\kappa$ B in several tumour cell lines and that the degree of tumour reduction is related to the inhibition of IKK (Fig. 6). It was suggested that the drug directly inhibited phosphorylation of I $\kappa$ B and thus the activation of NF- $\kappa$ B [63]. The survival of a neu-

roendocrine tumour cell line (TT: medullary thyroid carcinoma) and the tumour reduction of xenografted SCLC (NYH: small cell lung cancer) was found to be critically related to the activity of IKK [63, 64]. CHS828 was demonstrated to have potent anti-tumour effects also on xenografted neuroblastoma [65]. The concentration of CHS828 that caused apoptosis *in vitro* (1–10 nM) was in the same range as the IC<sub>50</sub> for IKK activity [63].

In our own studies, we have demonstrated a strong anti-tumour effect of CHS828, both *in vitro* and *in vivo*, on cell lines of three different neuroendocrine tumour types: ileal carcinoid (GOT1), medullary thyroid carcinoma (GOT2), and pancreatic carcinoid (BON). When xenotransplanted to nude mice, one weekly oral dose of CHS828 led to necrosis and complete tumour regression of both GOT1 and GOT2 within 2–3 weeks (250 mg/kg); a complete regression of BON tumours was also achieved, with no observed adverse effects [66] (Fig. 7).

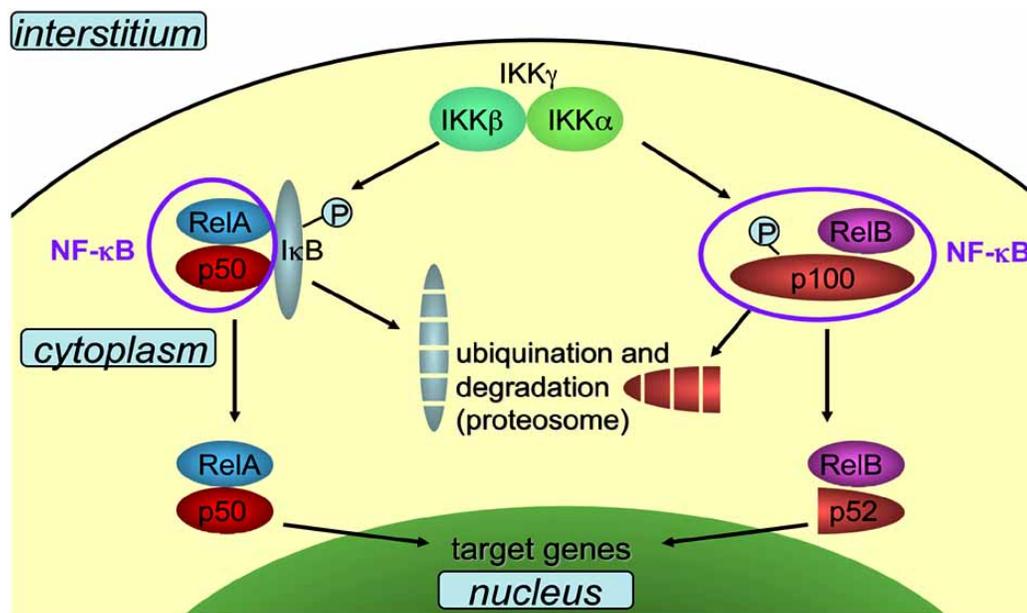


Fig. (5). Nuclear factor- $\kappa$ B signalling and interaction with the proteasome.

Nuclear factor- $\kappa$ B (NF- $\kappa$ B) is a group of transcription factors that mediate anti-apoptotic and pro-survival signals. NF- $\kappa$ B is constitutively activated in many tumour types. Inactive NF- $\kappa$ B is constrained in the cytoplasm by a binding protein, I $\kappa$ B (inhibitor of NF- $\kappa$ B). Upon stimulation, IKK (I $\kappa$ B kinase) phosphorylates I $\kappa$ B. This leads to ubiquitin-dependent degradation of I $\kappa$ B and release of the NF- $\kappa$ B heterodimer (i.e. RelA/p50), allowing it to translocate to the nucleus.

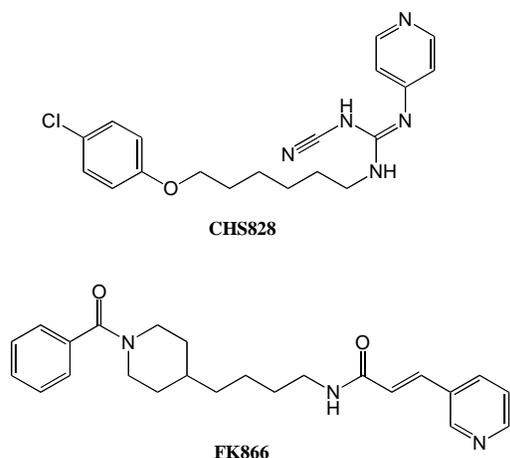


Fig. (6). Inhibitors of PARP and NAMPT.

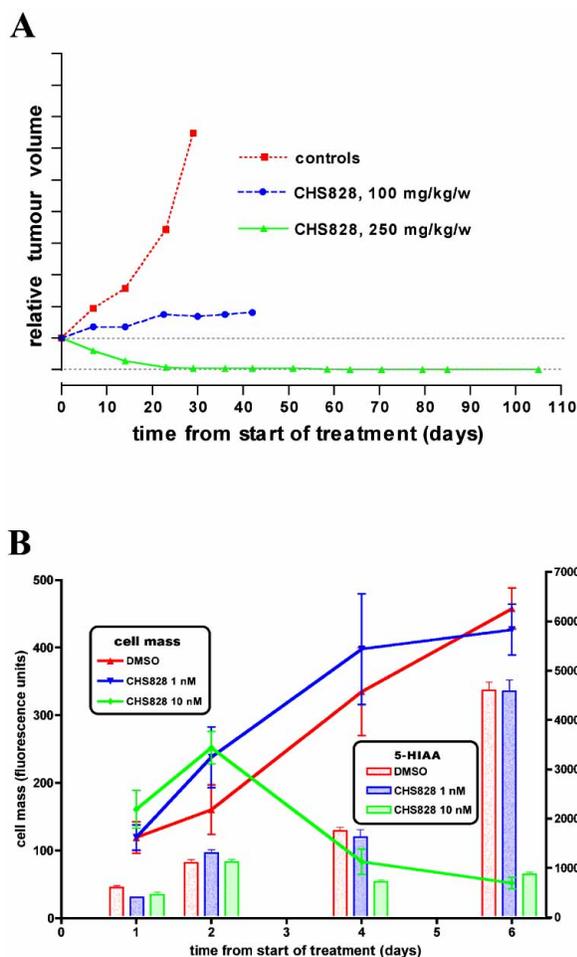


Fig. (7). Anti-tumour effects of CHS828.

(A) CHS828 was administered to nude mice xenografted with human midgut carcinoid (GOT1) once weekly by oral gavage: 100 mg/kg (days 0–42) or 250 mg/kg (days 0–57). The higher dose resulted in total tumour regression. (B) CHS828 was administered to the midgut carcinoid cell line GOT1 *in vitro* (0 nM (control), 1 nM, or 10 nM for 3 days). There was pronounced inhibition of cell growth at the higher dose, paralleled by depletion of intracellular serotonin.

An alternative mechanism of action for CHS828 involves NAD<sup>+</sup> metabolism [67]. Nicotinamide adenine dinucleotide (NAD<sup>+</sup>) is an essential co-enzyme in cellular redox reactions, e.g.

generation of ATP, but is also an important substrate in ADP ribosylation. Poly(ADP-ribose) polymerase 1 (PARP1) is an evolutionarily well-conserved nuclear enzyme with the key function of signalling genotoxic stress/DNA repair. The PARP family of enzymes includes 18 members, of which PARP1 and PARP2 are the only ones activated by DNA damage. Upon binding to DNA strand breaks, PARP1 metabolizes NAD<sup>+</sup> into branched polymers of ADP-ribose, which are transferred to nuclear proteins including PARP1 itself [68, 69]. Poly(ADP-ribosylation) has been proposed to function in genome repair by facilitating the opening of the condensed structure of chromatin required for the recruitment of the repairing enzymes. PARP1 activation thus consumes NAD<sup>+</sup>, which is cleaved to nicotinamide and ADP-ribose. NAD<sup>+</sup> can be resynthesized via the so-called “salvage pathway”. Thus, nicotinamide is first converted to nicotinamide mononucleotide (NMN) by the rate-limiting enzyme nicotinamide phosphoribosyltransferase (NAMPT) and then to NAD<sup>+</sup> by nicotinamide mononucleotide adenylyltransferase (NMNAT) [70]. Overactivation of PARP1 by massive DNA damage, e.g. radiotherapy, leads to depletion of NAD<sup>+</sup>/ATP energy stores and results in cellular death [71]. Tumour cells have high energy demands and PARP1 is constitutively activated due to DNA damage, which is the background to the new principle of cancer treatment by inhibition of NAD<sup>+</sup> synthesis [72]. Olesen and collaborators found that CHS828 reduced the intracellular levels of NAD<sup>+</sup> and impaired its re-synthesis to a similar degree to a structurally unrelated NAMPT inhibitor, FK866, and also demonstrated cross-resistance with this drug [67] (Fig. 6). CHS828 and FK866 were thus suggested to inhibit the NAD<sup>+</sup> salvage pathway [67] (Fig. 8). Watson and collaborators have recently confirmed that the mechanism of action of CHS828 is through potent and specific inhibition of NAMPT [73]. *In vitro* studies revealed that both NAD<sup>+</sup> and ATP were depleted by treatment with CHS828, but the NAD<sup>+</sup> depletion preceded that of ATP. Furthermore, nicotinic acid protected against the cytotoxicity caused by CHS828 (presumably via the Preiss-Handler pathway, an alternative salvage pathway of NAD<sup>+</sup> regeneration) (Fig. 8). The results indicated that the observed inhibition of IKK and NF-κB activation upon exposure to CHS828 is secondary to the NAD<sup>+</sup> depletion. Since GOT1 cells are sensitive to CHS828 both *in vitro* and as xenografts, the effects of CHS828 have to be compared with that of other NAMPT inhibitors in the same experimental models (Fig. 7).

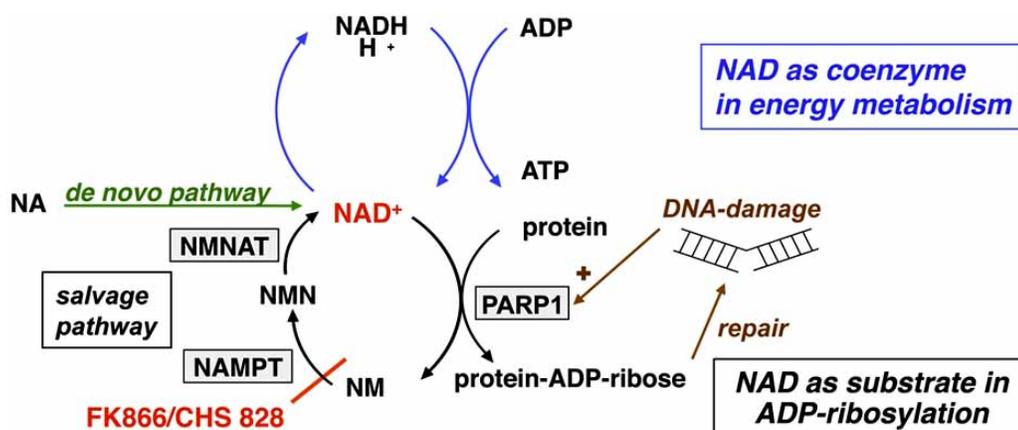
At present, 104 patients in total have received FK866, or CHS828, as monotherapy in phase-I trials [74]. Adverse effects with thrombocytopenia and gastrointestinal toxicity were significant and no tumour regression was seen in any of the five published studies. It has therefore been suggested that if NAD<sup>+</sup>-depleting drugs are to be used clinically, it will probably be in combination with therapies causing DNA damage and NAD<sup>+</sup> depletion [74].

### Targeting of the Proteasome

The 26S proteasome complex degrades proteins tagged with ubiquitin, and this mechanism is referred to as the ubiquitin-proteasome pathway (UPP). This non-lysosomal proteolytic system thus destroys proteins that are of importance for the cell cycle, apoptosis, and NF-κB activation (Fig. 5), as well as mutant proteins [75]. Inhibition of UPP is therefore an interesting target for cancer therapy. *Bortezomib* inhibits active threonine sites of the proteasome and has been tested on neuroendocrine tumour cells [76] (Fig. 9). In one small study of patients with carcinoids and EPT, disease stabilization was seen; neuropathy occurred in one third [77]. In studies on neuroendocrine tumour cells, the combination of histone deacetylase (HDAC) inhibition (see below) and bortezomib had marked anti-proliferative activity and induced apoptosis [78].

### Targeted Radionuclide Therapy

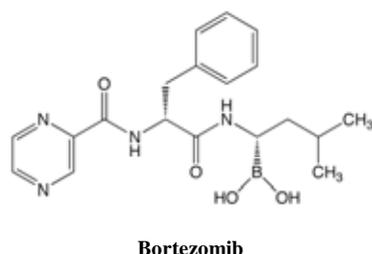
It is generally assumed that carcinoid tumours are relatively radioresistant, and that external radiation therapy is only indicated for palliation or for regionally advanced or metastatic disease [79-



**Fig. (8).** Interaction between NAD<sup>+</sup> and ATP production

NAD<sup>+</sup> is essential for enzymatic redox reactions involved in cellular metabolism, including ATP production. It is also a substrate of poly(ADP) ribosylation, a reaction catalyzed by PARP1 (poly(ADP-ribose) polymerase), which is activated by DNA strand breaks. This reaction consumes NAD<sup>+</sup>, which is cleaved to nicotinamide (NM) and ADP-ribose. NAD<sup>+</sup> can be re-synthesized either from exogenously administered nicotinic acid (niacin, NA) or from NM via the salvage pathway. Here, NM is first converted to nicotinamide mononucleotide (NMN) by nicotinamide phosphoribosyltransferase (NAMPT) and then to NAD<sup>+</sup> by nicotinamide mononucleotide adenylyltransferase (NMNAT). CHS828 and FK866 inhibit the NAD<sup>+</sup> salvage pathway as indicated.

84]. The assumption has persisted, although published studies on the clinical effects of external radiation therapy have actually revealed partial or complete remission in 25-80% of the patients after a radiation dose of 20–50 Gy to the tumour tissue [85-88].



**Bortezomib**

**Fig. (9).** Inhibitor of the proteasome.

Despite the assumed radioresistance of carcinoid tumours, SSTR-mediated radiation therapy has been introduced for neuroendocrine tumours, showing both promising effects and moderate toxicity [12, 89]. We have presented successful therapeutic results of <sup>177</sup>Lu-[DOTA<sup>0</sup>, Tyr<sup>3</sup>]-octreotate (<sup>177</sup>Lu-octreotate) in nude mice carrying GOT1 tumours [90] (Fig. 1). After intravenous injection of <sup>177</sup>Lu-octreotate, a rapid dose-dependent tumour regression was obtained (Fig. 10). Complete remission (>99% reduction of tumour volume) was obtained in 33% of the mice at 15 MBq, and in 100% at 30 MBq—corresponding to an absorbed dose of 60 and 120 Gy, respectively. Increased apoptosis was found one and three days after administration. Already 7 days after injection, the number of tumour cells was significantly reduced, and the necrosis and oedema demonstrated at day 3 were now replaced by fibrosis (Fig. 10).

We have previously found increased uptake of <sup>177</sup>Lu-octreotate in SCLC cells after irradiation *in vitro*, probably due to up-regulation of SSTR expression as demonstrated by increased SSTR2 mRNA expression [91]. These results were verified *in vivo* in nude mice xenografted with GOT1 [92]. After low amounts of <sup>177</sup>Lu-octreotate, a twofold higher concentration of subsequently given <sup>111</sup>In-octreotide was obtained in the tumour tissue. These results may be useful in optimizing therapy using radiolabelled somatostatin analogues. By choosing optimal time schedule and dosage, enhanced therapeutic effects might be obtained without increased side effects.

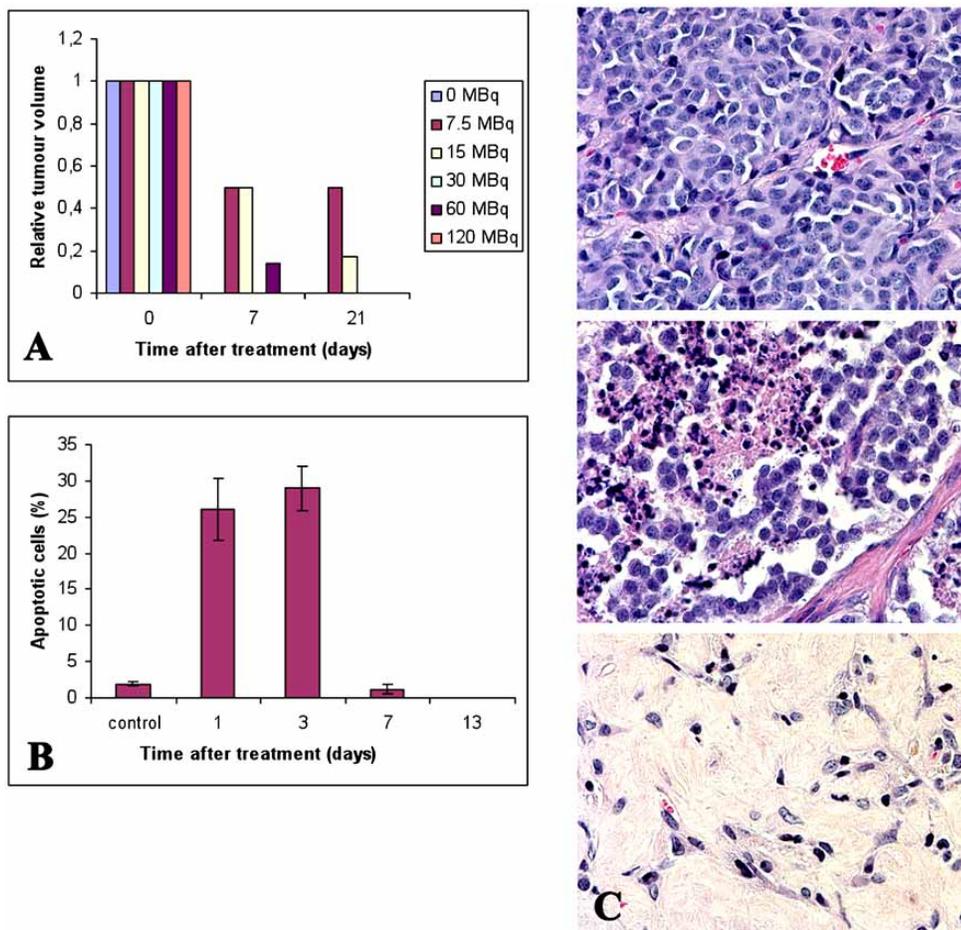
### Apoptotic Pathway

Ionizing radiation induces DNA damage, which is followed by a number of cellular responses, including DNA repair, cell cycle arrest, autophagy, mitotic catastrophe, necrosis, and apoptosis [93, 94]. Our understanding of the molecular mechanisms underlying the response to DNA damage is limited. Recent studies indicate that radiation-induced cell death can be divided into several distinct pathways according to the time course and the position in the cell-cycle; the apoptotic response plays a key role in almost every mode of cell death [94]. The tendency to undergo apoptosis varies between different cell types. Pre-mitotic apoptosis is rapid and is associated with a prompt activation of caspase 3, a key enzyme of intracellular signalling of apoptosis, while post-mitotic apoptosis is delayed until after cell division, and does not require rapid activation of caspase 3, but is associated with down-regulation of anti-apoptotic genes such as *MAPK* and *BCL2* [94]. Furthermore, radiation induces apoptosis both via the intrinsic mitochondrion-mediated and extrinsic death-receptor-mediated pathways. Apoptosis can thus be initiated in different cell compartments, e.g. the nucleus, the cytoplasm, or the plasma membrane [93]. There is evidence to suggest that radiation may induce biological responses via extranuclear targets and extracellular events [95-98].

Radiation-induced effects may vary in different types of tumour cells, but also between different types of normal cells due to the status of the signalling pathways in the cell. We have clearly demonstrated that irradiation by <sup>177</sup>Lu-octreotate induces apoptosis in GOT1 tumours in nude mice with a maximum at day 1 and 3 after administration of 30 MBq <sup>177</sup>Lu-octreotate [90] (Fig. 10).

### Bystander Effect

Radiation-induced bystander effects have been demonstrated for a variety of biological endpoints in both human and animal cell lines, whereby non-irradiated cells are affected by irradiated cells. The mechanisms behind this phenomenon are incompletely known [95, 96]. Mechanisms that have been suggested are communication via gap junctions, Ca<sup>++</sup> channels, cytokine or growth factor receptors, and small soluble mediators (reactive oxygen species (ROS), e.g. NO). The various signal transduction pathways involve the MAPK superfamily, i.e. ERK 1/2, c-Jun N-terminal kinase (JNK), and p38 kinase. Other mechanisms include the cyclooxygenase-2 (COX-2) signalling cascade, which can modulate cellular inflammation and genomic instability, IGF signalling pathways via IGFBP-3, and NAD(P)H oxidase, leading to long-lasting ROS pro-



**Fig. (10).** Anti-tumour effects of <sup>177</sup>Lu-octreotate

(A) <sup>177</sup>Lu-octreotate was administered to nude mice xenografted with human midgut carcinoid (GOT1); 7.5–120 MBq, resulted in dose-related tumour regression. (B) Relative number of apoptotic cells 1–13 days after treatment. (C) Micrographs of GOT1 cell xenografts in nude mice 0 (top), 7 (middle), and 13 days (bottom) after administration of 30 MBq <sup>177</sup>Lu-octreotate.

duction [96]. The *COX-2* gene is expressed in many carcinoids, and high expression of its protein in primary midgut carcinoids may be associated with a negative prognosis [99].

The importance of bystander effects in radiation therapy has been discussed, and they are potentially beneficial, especially in radionuclide therapy where more tumour cells could be affected than those targeted. The unexpectedly rapid GOT1 tumour cell reduction in our experimental studies might reflect a bystander effect (Fig. 10).

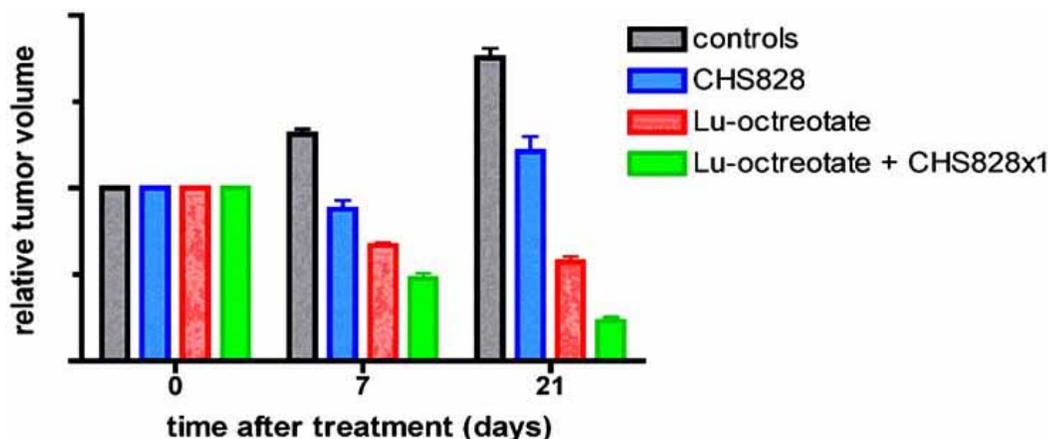
#### Interaction between Radiation and the Apoptotic Pathway

Anti-tumour agents that restore apoptosis signalling in cancer cells may induce tumour regression and improve conventional treatment such as radiation therapy. To enhance therapeutic effects, there is probably a need to combine radiation with one or several pro-apoptotic or anti-survival stimuli. In many tumours, including carcinoids, the apoptotic signalling is disrupted or reduced [58]. The relative radio-resistance might be due to such a resistance to apoptosis. One important way to enhance the cytotoxic effect of radiation therapy is by modulation of the apoptotic response (inhibition of anti-apoptotic signals) selectively in tumour cells by interventions involving specific signal transduction pathways (radio-sensitization).

There are several strategies to increase apoptotic signalling or reduce anti-apoptotic signalling in tumour cells, e.g. agonistic

monoclonal antibodies specific for death receptors, monoclonal antibodies competing with growth receptors, TKIs, small-molecule inhibitors to block e.g. Bcl-2 proteins, anti-sense oligonucleotides, or siRNA [100]. If such techniques are used, they should be active only on cancer cells without having any effect on normal cells. In many tumour types, several strategies have been suggested and tested both experimentally and in pilot clinical trials [100].

Carcinoids express *BCL2* and *MYC* to a high degree, while *p53* is seldom over-expressed or mutated [101, 102]. High levels of Bcl-2 protein expression may thus be linked to the intrinsic resistance to chemotherapy and radiation therapy [101]. Bcl-2 inhibits apoptosis, and inhibition of this protein might stimulate apoptosis and increase the radio-sensitivity of carcinoids. As mentioned above, nude mice xenografted subcutaneously with GOT1 tumours were found to be curable with high doses of <sup>177</sup>Lu-octreotate or CHS828 (30 MBq intravenously and 250 mg/kg/week orally, respectively) [66, 90]. In a recent experimental study on xenografted nude mice, we studied CHS828 as a radio-sensitizer. Much lower doses of <sup>177</sup>Lu-octreotate or CHS828 (7.5 MBq and 100 mg/kg/week, respectively) resulted in partial regression of tumours, while the combination of the two therapies resulted in complete tumour regression in almost all the animals (Fig. 11). As CHS828 inhibits the regeneration of NAD<sup>+</sup> and since there is a synergistic effect of CHS828 and radiation therapy, PARP1 activation may be involved in the death mechanism induced by radiation under these conditions.

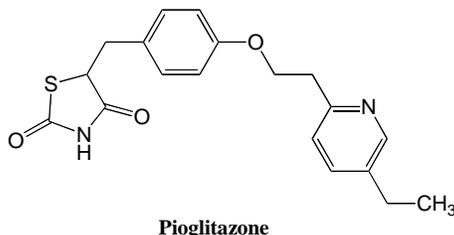


**Fig. (11).** Radio-sensitizing effects of CHS828 on carcinoid tumour cells

Nude mice xenografted with human midgut carcinoid GOT1 were treated with  $^{177}\text{Lu}$ -octreotate (7.5 MBq intraperitoneally) with a single dose of CHS828 (100 mg/kg per os) or with a combination with these two treatments; the combined treatment resulted in enhanced therapeutic effect.

### Death Receptor Signalling

The death receptors belong to the tumour necrosis factor (TNF) receptor superfamily, which is characterized by cysteine-rich extracellular domains and a cytoplasmic death domain that serves as the recognition point for the apoptotic machinery. Much interest has been focused on death receptor signalling, especially regarding TRAILR1 and TRAILR2. TRAIL, the natural ligand of TRAILR1 and -2, preferentially kills tumour cells but has no effect on normal cells. However, TRAIL has a short half-life *in vivo* [103-106], which makes it a less attractive alternative for therapy. Instead, agonistic TRAILR antibodies and recombinant ligands have been developed. TRAIL resistance may also be due to increased expression of anti-apoptotic proteins, e.g. Bcl-2 and c-FLIP [107]. In gastrointestinal tumours, specific TRAIL-induced signalling can be obtained from proteasome or HDAC inhibitors with no response in normal cells [100, 108, 109] (see below). TRAIL-induced apoptosis has been studied in carcinoid cells *in vitro*, and a peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ) agonist *pioglitazone* (Fig. 12) inhibits carcinoid cell growth and promotes TRAIL-induced apoptosis [110]. Since ionizing radiation has been shown to up-regulate the *TRAILR1* and -2 genes and/or the *TRAIL* gene through p53- and p63-dependent mechanisms causing an autocrine or paracrine type of apoptosis [103], a combination of radiation and a TRAIL analogue and/or a PPAR- $\gamma$  analogue might be valuable as therapy.

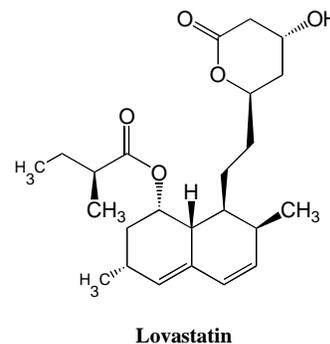


**Fig. (12).** Agonist of PPAR- $\gamma$ .

By activating SAPK/JNK, the Fas death receptor was expressed and shown to facilitate apoptosis in irradiated leukemia cells [111]. Post-irradiation events such as membrane damage (by anti-oxidant inhibition of lipid peroxidation) further induce caspase 3-dependent apoptosis, with no change in expression of the *BCL2* and *BAX* genes [112]. The extent to which Fas receptors are expressed in carcinoids is, however, unclear.

### Possible Interaction Between Radiation and Other Signalling Pathways

Over-expression of RAS has been shown to increase resistance to radiation [113, 114]. Radio-sensitivity may thus be manipulated in pathways upstream or downstream from RAS. One method of increasing the radiosensitivity is by inhibition of RAS activation, e.g. by using *lovastatin* and farnesyl-transferase inhibitors (Fig. 13) to block the processing of RAS [115, 116]. *EGFR* expression and AKT phosphorylation are also associated with the response to radiation, and inhibition of EGFR, RAS, PI3K, and AKT has been shown to increase the radio-sensitivity of cancer cell lines. PI3K is a mediator of RAS-induced radiation resistance. EGFR, RAS, and PTEN can also regulate the PI3K pathway. Molecular-based radiosensitization could thus be directed against signals that are common to these pathways [113].



**Fig. (13).** Inhibitor of RAS activation.

As mentioned above, neuroendocrine tumours frequently express VEGF, which is correlated to tumour progression [36]. VEGF leads to increased angiogenesis and higher oxygen concentration. Higher oxygen levels in the tumour tissue enhance the cytotoxic effect of radiation (oxygen effect). On the other hand, radiation therapy also increases expression of VEGF, due to radiation stress of the tumour, and may contribute to resistance to treatment through higher cell survival and proliferation. Blocking of the radiation-mediated increase in VEGF with anti-VEGF therapy could therefore increase the destruction of tumour cells and produce additive anti-tumour effects to that of radiation alone. Clinical trials designed to address this issue are in progress.

### Epigenetic Therapy and Targeting of Notch Signalling

Changes in gene expression due to mechanisms other than altered DNA sequences are termed epigenetic changes. During tumorigenesis, global hypo-methylation and promoter hyper-methylation are commonly observed. DNA methylation of cytosines occurs at CpG sites (4% of the genome), i.e. CG dinucleotide clusters in small sequences of DNA mostly in the gene promoter regions. The methylation is a covalent addition of a methyl group to the 5'-carbon of the cytosine ring, which results in 5'-methylcytosine. DNA hypermethylation contributes to tumour development by predisposing to mutations at methylated CpG sequences and by silencing of specific genes, repressing gene transcription by inhibiting the binding of transcription factors, or by recruiting methyl CpG-binding proteins [117]. Several genes are hyper-methylated in ileal carcinoids, e.g. *RUNX3*, *O<sup>6</sup>-MGMT*, *RASSF1A*, *p14*, and *CTNNB1*. Some of these are suppressor genes associated with advanced disease [118-120]. The cytidine analogues 5-azacytidine and 5-azadeoxycytidine are demethylating agents currently being tested as treatment for patients with different tumour types. Global hypo-methylation influences the repetitive interspersed nuclear elements (45% of the genome), which normally protects against harmful retroviral sequences. Loss of this function may lead to loss of heterozygosity and gene rearrangements [117]. Global hypo-methylation is more common in carcinoids than in EPT, and is associated with features such as loss of chromosome 18, methylation of *RASSF1A*, and lymph node metastasis [121].

Another epigenetic mechanism is modification of histones, which are proteins associated with DNA in the chromatin complex. As part of gene regulation the histones are acetylated, or deacetylated, on lysine residues in the N-terminal tail. These reactions are catalyzed by enzymes with histone acetyltransferase (HAT) or histone deacetylase (HDAC) activity. The removal of acetyl groups increases the positive charge of the histone tails, which promotes binding between the nucleosome and the negatively charged phosphate groups of the DNA backbone. The increased DNA binding condenses the chromatin structure and prevents transcription of genes. Down-regulation of tumour suppressor genes, evasion from apoptosis, and reduced differentiation are hallmarks of cancer that can be due to abnormal epigenetic control caused by de-regulated HDAC activity. HDAC inhibitors are emerging as anti-cancer drugs with the capacity to prevent silencing of genes and to reverse crucial steps in tumorigenesis. These inhibitors can thus induce apoptosis and differentiation, suppress proliferation and angiogenesis, and enhance the host immune system [122]. However, HDAC can also modulate the acetylation of cytoplasmic proteins, e.g. p53, the STAT family, E2F, tubulin, and heat shock protein 90 (HSP90), which are involved in a variety of cellular mechanisms. Thus, HDAC inhibitors can act through both epigenetic and non-epigenetic mechanisms. HDAC inhibitors have a broad molecular range of targets and new inhibitors are continuously under development.

Notch signalling is an important developmental pathway and plays a central role in stem cell maintenance, cell fate decisions, and differentiation. The Notch protein is a cell membrane receptor that becomes activated upon ligand binding to the extracellular domain. Induced proteolytic cleavage by  $\gamma$ -secretase releases the intracellular domain of the Notch receptor, which enters the cell nucleus, binds to DNA, and alters gene expression. Depending on cellular context, Notch can act either as a tumour suppressor or as an oncogene. In hepatocellular carcinoma and in neuroendocrine tumours, e.g. SCLC, medullary thyroid cancer, and gastrointestinal carcinoids, Notch signalling is very low or absent and is associated with growth suppression. In these tumour types, Notch is considered to be a tumour suppressor [123-126]. Notch signalling is a negative regulator of the expression of achaete-scute complex homolog 1 (ASCL1); the ASCL1 protein is expressed in both progenitor cells and neuroendocrine tumour cells.

The amyloid precursor-like protein 1 (APLP1) belongs to the amyloid precursor protein (APP) family, which, like Notch, are activated by  $\gamma$ -secretase. There is documented cross-talk between the APP proteins and Notch. One mechanism occurs through interactions between the cleaved fragment of APP and cytoplasmic inhibitors of the Notch receptor [127]. The second way is through interaction between APP and Notch by heterodimerization [128]. Gastrointestinal neuroendocrine tumours and the GOT1 cell line have elevated expression of APLP1 in comparison with non-neuroendocrine gastrointestinal tumours. This finding raises the possibility that signals from the APP and Notch protein families may interact and regulate the differentiation of neuroendocrine cells. The increased expression of APLP1 in hepatic metastases in comparison to that in primary ileal carcinoids indicates that it may have a role in invasiveness and spread [129].

The HDAC inhibitor *valproic acid* (VPA) is a branched fatty acid with an established safety profile as an anti-epilepsy drug (Fig. 14). Stockhausen and collaborators showed that VPA caused elevated levels of Notch1 in neuroblastoma cells [130]. Recent work on BON cells and pulmonary carcinoid cells has shown that the drug causes cell cycle arrest in G1 and suppresses the synthesis of CgA. These effects were found to be associated with activation of Notch1 (increased full-length Notch1 and its active intracellular domain). Reporter assays verified that VPA-induced Notch1 was functionally active (with binding to the centromere-binding factor and the ASCL1 promoter); siRNA against Notch1 inhibited the drug-induced effects on proliferation [131].

HDAC inhibitors alone, or in combination with other anti-cancer drugs, are currently in clinical trials for a wide range of tumour types. However, until now clinical studies on patients with ileal carcinoids using HDAC inhibitors have been very limited. A phase-I study has been conducted using *entinostat* (*MS-275*) in treatment of patients with advanced solid tumours including carcinoids [132] (Fig. 14). A phase-II study using *romidepsin* (*depsipeptide/FK228*) in patients with locally advanced or metastatic neuroendocrine tumours was terminated due to an unexpected high number of serious cardiac adverse events [133] (Fig. 14).

### Targeting of Chemokine Receptors

Directional migration of cells during embryonic development, during tissue repair, and during the process of homing of hematopoietic stem cells to distinct niches is regulated by chemokines. Chemokines bind and activate specific G protein-coupled chemokine receptors. The chemokine receptors are divided into different families according to structure: the CXC-, CC-, CX3C-, and XC- chemokine receptors. The chemokine CXCL12 (stromal cell-derived factor 1/SDF1) activates the CXC chemokine receptor CXCR4. *In vitro*, CXCR4-expressing cells migrate towards a gradient of CXCL12. *In vivo*, CXCL12 is secreted by mesenchymal fibroblasts, osteoblasts, and endothelial cells. There is substantial evidence to suggest that CXCL12-CXCR4-mediated migration is crucial for the metastatic process and tissue-specific spread of breast and prostate cancer [134-138]. To date, three types of neuroendocrine tumours have been shown to express CXCR4: SCLC, neuroblastoma, and ileal carcinoids [139-141]. In ileal carcinoid cells (GOT1), we recently found that stimulation of CXCR4 by its ligand CXCL12 markedly activates the ERK1/2 signalling pathway and promotes cell migration under hypoxic conditions [139]; CXCR4 is thus a potential marker of tumour stem cells and hypoxic tumour cells [142]. Pre-clinical testing of a number of CXCR4-antagonistic peptides or small molecules, and antibodies to CXCR4 or CXCL12, has been conducted in mouse tumour models with promising results [138, 143-147]. Some of these small molecules are currently being investigated in acute myeloid leukaemia and multiple myeloma, but not in neuroendocrine tumours [148].

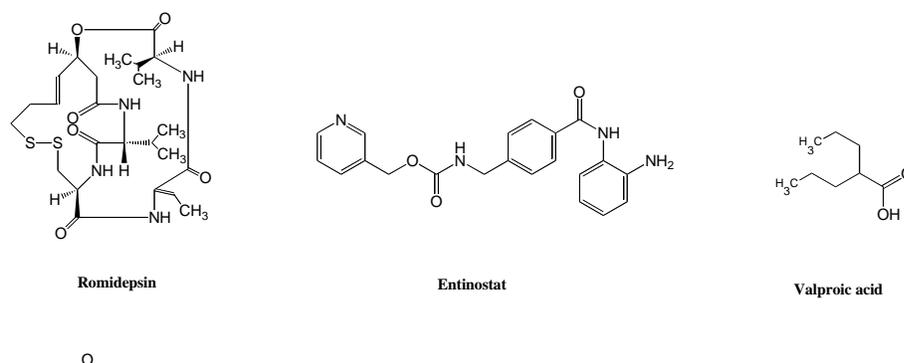


Fig. (14). Inhibitors of HDAC.

### Targeting of IGF1R, EGFR, and TGF Signalling

Insulin-like growth factor 1 (IGF1) signalling has emerged as an important mechanism in the development and progression of cancer [149, 150]. A key regulator in this signalling pathway is IGF1R, which is widely expressed in malignant tumours including neuroendocrine tumours [151]. Over-expression of IGF1R in tumour cells causes increased proliferation and resistance to chemotherapy and apoptosis. IGF1R is a tetrameric membrane protein with an extracellular ligand-binding domain, a transmembrane domain, and an intracellular tyrosine kinase domain. Activation of IGF1R is achieved by phosphorylation of the tyrosine kinase, which in turn leads to activation of the AKT and MAPK signalling pathways. Targeting of IGF1 signalling has therefore emerged as a promising therapeutic strategy [149].

The role of IGF1 signalling has been investigated in the pancreatic carcinoid cell line BON; BON cells secrete both IGF1 and IGFBP2 and they express functional IGF1R [152]. Exogenously supplied IGF1 was found to stimulate BON cells to secrete CgA and to proliferate. It has been suggested that endogenous IGF1 has a role as an autocrine regulator of carcinoid tumour cell growth. The effects of IGF1 stimulation were shown to be mediated by activation of PI3 kinase, p70<sup>s6k</sup>, and MAPK/ERK [152], and also by activation of AKT and mTOR [153]. Inhibition of IGF1R phosphorylation by *NVP-AEW541* was found to cause cell cycle arrest, activation of caspase 3, and apoptosis in BON cells [154] (Fig. 15). Forced activation of the RAF1 signalling pathway in BON cells was also shown to interfere with MEK1/2 and IGF1 signalling, causing reduced production of CgA. However, to date IGF1 signalling has not been evaluated in the small intestinal carcinoid cell lines KRJ-I and GOT1, although an autocrine role for IGF1 has been demonstrated in primary cell cultures of ileal carcinoids [155]. At present, there are no published clinical studies on the effectiveness of IGF1R inhibition in ileal carcinoids.

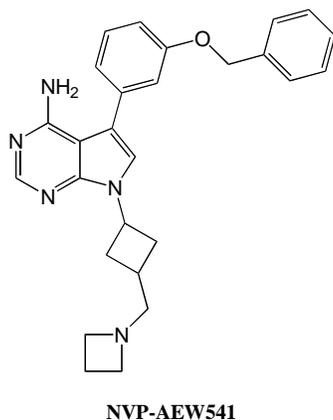


Fig. (15). Inhibitor of IGF1R.

The epidermal growth factor receptor (EGFR) is frequently activated in malignant tumours e.g. colorectal carcinoma, NSCLC, head and neck squamous cell carcinoma, and pancreatic cancer [156]. Activated EGFR has been successfully targeted in these tumours using small molecules that inhibit RET or monoclonal antibodies that interfere with receptor dimerization. Constitutive activation of the EGFR receptor in tumours is the result of gene amplification or mutations in the extracellular part of the receptor, or in the tyrosine kinase domain. Mutations in EGFR strongly influence the responsiveness to TKI, while mutations in downstream effector molecules, e.g. *KRAS*, *BRAF*, *PIK3AC*, or *PTEN* confer resistance to EGFR-targeted therapy. The EGFR signalling pathway has been investigated in pancreatic and ileal carcinoid cells (BON and KRJ-I cells, respectively). EGFRs were demonstrated in BON cells, and exogenous administration of EGF or TGF $\alpha$  induced a proliferative response [53, 157, 158]. Treatment with the EGFR inhibitors *gefitinib* or *AG1478* induced an anti-proliferative response due to cell cycle arrest and apoptosis. In KRJ-I cells, however, TGF $\alpha$  induced cell proliferation whereas EGF did not [53]. Treatment with gefitinib, on the other hand, inhibited proliferation of KRJ-I cells [159]. Furthermore, studies on primary cell cultures from ileal carcinoids have found an autocrine stimulatory effect of TGF $\alpha$  that could be blocked by neutralizing EGFR antibodies [160]. The experimental data thus clearly indicate that EGFR signalling is of major importance in carcinoid tumour cells. Clinical studies on patients with carcinoid tumours have demonstrated disease stabilization, or even regression of tumours, after EGFR inhibition with gefitinib as monotherapy [18].

The transforming growth factor  $\beta$  (TGF $\beta$ ) family has 33 members; these control developmental processes as well as adult homeostasis. TGF $\beta$ s regulate tissue homeostasis by their ability to induce cell cycle arrest and apoptosis, and to preserve genomic stability. In malignant tumours, the TGF $\beta$  signalling pathway is frequently perturbed, causing resistance to the cytostatic activities of TGF $\beta$ . The oncogenic properties of TGF $\beta$  are poorly understood, but include regulation of the tumour microenvironment, of angiogenesis, of the epithelial-mesenchymal transition, and of metastasis formation [161]. TGF $\beta$  ligands signal via receptor serine/threonine kinases (TGF $\beta$ R) that undergo phosphorylation upon ligand binding and activate intracellular SMAD effectors. Oligomeric SMAD complexes translocate to the nucleus, associate with chromatin, and regulate gene transcription [162]. The TGF $\beta$  signalling pathway has been investigated in pancreatic (BON) and ileal (KRJ-I) carcinoid cells. Surprisingly, BON cells and KRJ-I cells showed opposite proliferative responses to exogenous TGF $\beta$ . BON cell growth was inhibited, while KRJ-I cell growth was stimulated after administration of TGF $\beta$  [53, 163]. Stimulation of BON cells with TGF $\beta$  induced G1 cell cycle arrest, reduced expression of *MYC*, and increased expression of *p21<sup>WAF1/CIP1</sup>*. Furthermore, TGF $\beta$  was found to be secreted from tumour cells, suggesting that there is an autocrine growth inhibition by TGF $\beta$  [164]. The relationship be-

tween TGF $\beta$  and somatostatin signalling has been investigated in BON cells [165]. Treatment with TGF $\beta$  caused increased expression of somatostatin and SSTR2 in tumour cells. Administration of the SSTR antagonist *cyclo-somatostatin* abolished the growth inhibition induced by TGF $\beta$ , indicating that the growth-inhibitory effect of TGF $\beta$  is mediated by SSTRs (Fig. 16). The TGF $\beta$  signalling pathway has also been characterized in KRJ-I cells. In these cells, TGF $\beta$  stimulation did not induce nuclear translocation of SMAD or growth inhibition; on the contrary, TGF $\beta$  induced cell proliferation [166]. TGF $\beta$ -stimulated growth was associated with SMAD2 phosphorylation, but with reduced SMAD4 expression and increased SMAD7 expression. The TGF $\beta$  target gene *p21<sup>WAF1/CIP1</sup>* was down-regulated, while *c-MYC* was up-regulated. These molecular changes together with phosphorylation of ERK1/2 partly explain the proliferative response of KRJ-I cells upon TGF $\beta$  stimulation. Identification of an aberrant TGF $\beta$  signalling pathway in ileal carcinoids may provide novel targets for therapy. Anti-TGF $\beta$  compounds are under development and have proven efficacy in pre-clinical studies [167]. However, there is still a lack of clinical studies on carcinoid tumours.

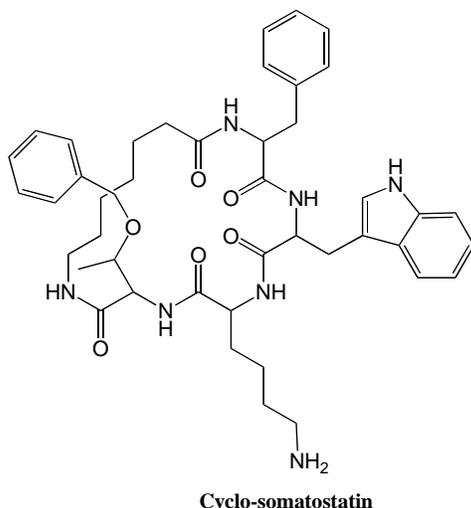


Fig. (16). SSTR antagonist.

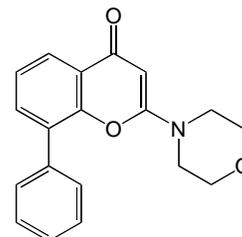
A number of additional growth factors/receptors have been identified and functionally characterized in carcinoid cells, including nerve growth factor receptor (NGFR) and connective tissue growth factor (CTGF) [53, 168]. The exact role of these factors and receptors in carcinoid growth regulation and metastasis remains to be clarified, as does their suitability as targets for anti-cancer therapy.

#### Targeting of PI3K-AKT Signalling

In several tumour types, including the neuroendocrine tumours such as medullary thyroid carcinoma, up-regulation of the PI3K-AKT signalling pathway can promote tumour growth and inhibit apoptosis [169]. AKT (protein kinase B) is the key effector; it must be phosphorylated at a threonine and serine site for full activity. This is carried out by PI3K and phosphate-dependent dehydrogenase-1. Phosphorylated AKT in turn controls several caspases and transcription factors of the Forkhead family. AKT occurs in three isoforms; of these, AKT1 is the main one, AKT2 occurs in insulin-responsive tissues, and AKT3 occurs in the central nervous system [170].

Studies on human pulmonary carcinoid cells (NCI-H727) subjected to blockade with *LY294002*, a well-characterized PI3K inhibitor, or subjected to silencing with AKT1 siRNA showed reduced growth and reduced expression of neuroendocrine markers (CgA and ASCL-1) but to different degrees (85% as opposed to 31%) [171] (Fig. 17). The study did not address whether anti-

proliferative or apoptotic mechanisms were activated. Previous studies on endocrine-related tumours have shown that AKT1 regulates migration and invasiveness in breast cancer [172] and neuroendocrine differentiation in prostate cancer [173]. Of special interest for therapy with <sup>177</sup>Lu-octreotate is that targeting of AKT1 enhances the radiation toxicity of human tumour cells by inhibiting the DNA-PK-dependent DNA double-strand break repair [174]. PI3K-AKT signalling has been implicated also in solid tumours and haematological malignancies, and clinical trials with PI3K-AKT inhibitors have been initiated. No experimental studies on gastrointestinal carcinoids have been reported.



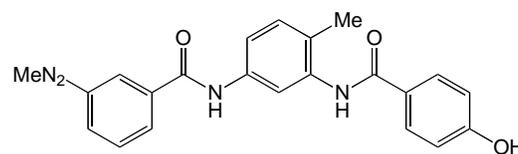
**LY294002**

Fig. (17). Inhibitor of PI3K-AKT.

#### Targeting of RAS/RAF1 Signalling

In this pathway, RAS activates RAF1, which is a serine/threonine kinase, resulting in phosphorylation of MEK 1/2 and ERK 1/2. The pathway can be constitutively activated by oncogenic RAS/RAF isoforms, or over-expression of RAS/RAF [175]. This pathway is commonly activated in gastrointestinal carcinomas, but mutations in RAS are unusual in intestinal carcinoids [176]. *BRAF* mutations are also rare in neuroendocrine tumours, but the activating protein RAP1 is frequently expressed [177]. In a recent report, RAF1/BRAF signalling was shown to be activated in neuroendocrine tumours, which makes this pathway a putative therapeutic target [178]. Notably, RAF1 activation in medullary thyroid cancer leads to reduced tumour growth and reduced levels of neuroendocrine tumour markers [125].

Pharmacological activation of RAF1 by *ZM336372* was found to induce progressive phosphorylation of RAF1, MEK 1/2, and ERK 1/2 in both human pulmonary carcinoid cells and BON (pancreatic carcinoid) cells, accompanied by reduced levels of CgA and the transcription factor ASCL1 [179] (Fig. 18). The treatment suppressed proliferation, possibly through induction of cell cycle inhibitors such as p18 and p21. The results must be interpreted with some caution, since the drug may phosphorylate proteins other than RAF 1 [180]. The basic helix-loop transcription factor ASCL1 was previously shown to be highly expressed in neuroendocrine tumour cells [181]. MASH-1 is the mouse homologue of ASCL1 and is important for foetal development of certain neurons, thyroid C-cells, and adrenal chromaffin cells [182-184]. Several of the studies cited indicate that ASCL1 controls the expression of neuroendocrine granule proteins, i.e. depletion of ASCL1 is associated with reduction of CgA and synaptophysin expression. Pharmacological RAF1 activation may therefore be a future therapeutic option with the potential to alleviate hormonal symptoms. No experimental studies have been performed on midgut carcinoid tumour cells.



**ZM336372**

Fig. (18). Activator of RAF1.

**CONCLUSION**

Targeting of growth factors and their receptors, angiogenic factors, and hormone receptors expressed on neuroendocrine tumour cells provides new therapeutic options. The signal transduction in carcinoid tumour cells is not fully understood, but experimental interference with specific pathways, e.g. mTOR, PI3K, RAS/RAF, and Notch, offers some promise aside from more general mechanisms such as inhibition of the proteasome and HDAC. The combination of several therapeutic strategies seems rational, since it gives tumour cells limited options for escape. In clinical studies, combination therapies (based on TKIs) have had acceptable toxicity. Radionuclide therapy via highly expressed SSTRs has been shown to be effective clinically in limited, but disseminated tumour disease. New knowledge gained concerning the apoptotic pathway, PARP inhibition, and death receptors could lead to potentiated radiotherapeutic effects.

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**ABBREVIATIONS**

ADP	= Adenosine diphosphate	CIP1	= Cyclin-dependent kinase inhibitor 1A, also known as p21, WAF1, or CDKN1A; a cell cycle inhibitor
AKT	= v-Akt murine thymoma viral oncogene homolog, a serine-threonine protein kinase, also known as protein kinase B	c-MYC	= A gene encoding for a transcription factor regulating expression of many genes
AKT1	= Involved in cellular survival pathways, a major factor in many cancers	CNDT2	= A human ileal (midgut) carcinoid cell line
AKT2	= Important signalling molecule in the insulin signalling pathway	CNS	= Central nervous system
AKT3	= Occurs predominantly in the central nervous system	COX-2	= Cyclooxygenase-2
AG1478	= An EGFR inhibitor	CpG	= Cytosine-phosphate-guanine dinucleotide sequence of DNA
AMG706	= A pan-VEGFR inhibitor, a nicotinamide	c-RAF	= A gene that codes for the protein kinase c-RAF; also called RAF1
APLP1	= Amyloid precursor-like protein 1	CTGF	= Connective tissue growth factor
APP	= Amyloid precursor protein	CTNNB1	= A gene coding for $\beta$ -catenin
ASCL1	= Achaete-scute complex homolog 1	CX3C	= A type of chemokine
ATP	= Adenosine triphosphate	XC	= A family of chemokines
BAX	= BCL2-associated X protein	CC	= A family of chemokines
BCL2	= A family of genes and proteins that are either pro- or anti-apoptotic	CXC	= A family of chemokines
bFGF	= Basic fibroblast growth factor	CXCR4	= Chemokine (C-X-C motif) receptor 4, also known as fusin
BON	= A human pancreatic carcinoid cell line	DNA	= Deoxyribonucleic acid
BRAF	= V-raf murine sarcoma viral oncogene homolog B1	DOTA	= 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid, a chelate
cAMP	= Cyclic adenosine monophosphate, a second messenger	E2F	= A group of genes that codes for a family of transcription factors, involved in cell cycle regulation and DNA synthesis
caspase	= Cysteine-aspartic acid protease, involved in, e.g., apoptosis, and necrosis	EGF	= Epithelial growth factor
CDKN1A	= Cyclin-dependent kinase inhibitor 1A, also known as p21, WAF1, or CIP1; a cell cycle inhibitor	EGFR	= Epithelial growth factor receptor
cFLIP	= FLICE (FADD-like IL-1 $\beta$ -converting enzyme) inhibitory protein, a protease-deficient caspase homologue	EPT	= Endocrine pancreatic tumour
CgA	= Chromogranin A	EORTC	= European Organisation for Research and Treatment of Cancer
CHS828	= A pyridyl cyanoguanidine with anti-tumour activity	ERK	= Extracellular signal-regulated kinase; also known as mitogen-activated protein kinase, MAPK
		ERK1	= Also known as MAPK3, an enzyme encoded by the <i>MAPK3</i> gene
		ERK2	= Also known as MAPK1, and p42MAPK, an enzyme encoded by the <i>MAPK1</i> gene
		FADD	= Fas-associated protein with death domain
		FasR	= Fas receptor, one of the death receptors
		FGFR1	= Fibroblast growth factor receptor 1, also known as basic fibroblast growth factor receptor 1, fms-related tyrosine kinase-2/Pfeiffer syndrome, and CD331
		FK228	= An HDAC inhibitor, also called depsipeptide
		FK866	= An NAMPT inhibitor
		FLT3	= Fms-like tyrosine kinase 3
		FOLFOX	= A chemotherapy regimen consisting of 5-fluorouracil, leucovorin, and oxaliplatin
		G1	= Gap 1, one of the cell cycle phases
		GIST	= Gastrointestinal stromal tumour
		GMX1778	= An inhibitor of nicotinamide phosphoribosyl-transferase (NAMPT)
		GOT1	= A human ileal (midgut) carcinoid cell line
		GOT2	= A human medullary thyroid carcinoma cell line
		Gy	= Gray, unit of absorbed dose

HIF-1	= Hypoxia-inducible factor 1	NO	= Nitric oxide
HSP90	= Heat shock protein 90	NSCLC	= Non-small cell lung carcinoma
HAT	= Histone Acetyltransferase	NVP-AEW541	= A selective IGF1R inhibitor
HDAC	= Histone Deacetylase	NYH	= A human small cell lung carcinoid cell line
IC <sub>50</sub>	= Half maximal inhibitory concentration	O <sup>6</sup> -MGMT	= O <sup>6</sup> -methylguanine-DNA methyltransferase, interacts with estrogen receptor alpha
IGF	= Insulin-like growth factor	p14	= The fusion-associated small transmembrane (FAST) protein
IGF1	= Insulin-like growth factor 1	p18	= Cyclin-dependent kinase 4 inhibitor C, a cell cycle inhibitor
IGF1R	= Insulin-like growth factor 1 receptor	p21	= Cyclin-dependent kinase inhibitor 1A, also known as WAF1, CIP1, or CDKN1A; a cell cycle inhibitor
IGFBP2	= Insulin-like growth factor binding protein 2	p38	= A class of mitogen-activated protein kinases involved in cell differentiation and apoptosis
IGFBP3	= Insulin-like growth factor binding protein 3	p50	= A mature subunit of NF-κB
IgG	= Immunoglobulin G	p53	= Protein 53, or tumour protein 53, is a tumour suppressor protein; also known as cellular tumour antigen p53, antigen NY-CO-13, phosphoprotein p53, transformation-related protein 53 (TRP53), or tumour suppressor p53
IκB	= Inhibitory protein of NF-κB	p63	= Tumour protein p63, also known as transformation-related protein 63, a member of the p53 family of transcription factors
IKK	= Inhibitory protein of NF-κB kinase	p70 <sup>s6k</sup>	= p70 ribosomal S6 protein kinase
In	= Indium	PARP	= Poly (ADP-ribose) polymerase; involved in, e.g., DNA repair and apoptosis
JNK	= C-Jun N-terminal kinase	PARP1	= Poly (ADP-ribose) polymerase 1
KIT	= A tyrosine kinase receptor, also known as C-kit or CD117	PARP2	= Poly (ADP-ribose) polymerase 2
KRJ-1	= A human ileal (midgut) carcinoid cell line	PDGF	= Platelet-derived growth factor
KRAS	= V-Ki-ras2 Kirsten rat sarcoma viral oncogene homolog	PDGFA	= PDGF isoform A
LAR	= Long acting release	PDGFB	= PDGF isoform B
Lu	= Lutetium	PDGFR	= Platelet-derived growth factor receptor, a receptor tyrosine kinase
LY294002	= A selective phosphatidylinositol 3-kinase (PI3K) inhibitor	PDGFRA	= PDGFR alpha isoform
MASH-1	= The mouse homologue of ASCL1	PI3K	= Phosphatidylinositol 3 kinase, involved in the PI3K/AKT signalling pathway
MAP	= Mitogen-activated protein	PIK3CA	= The catalytic subunit of PI3K
MAPK	= Mitogen-activated protein kinase	PIGF	= Placental growth factor
MEK1	= Dual-specificity mitogen-activated protein kinase kinase 1, also known as ERK activator kinase 1 or MAPK/ERK kinase 1	PIGF1	= Placental growth factor 1
MEK2	= Dual-specificity mitogen-activated protein kinase kinase 2, also known as ERK activator kinase 2, or MAPK/ERK kinase 2	PIGF2	= Placental growth factor 2
MGMT	= Methylguanine-DNA methyltransferase	PPAR-γ	= Peroxisome proliferator-activated receptor gamma
mRNA	= Messenger ribonucleic acid	PTEN	= Phosphatase and tensin; acts as a tumour suppressor gene
MS275	= Entinostat, an HDAC inhibitor	RAF1	= A serine/threonine kinase, also known as RAF or c-RAF
mTOR	= Mammalian target of rapamycin	RAP1	= Ras-proximate-1, a small GTPase involved in signal transduction
MYC	= V-myc myelomatosis viral oncogene homolog, a transcription factor regulating cell cycle progression and apoptosis	RAS	= A family of genes encoding small GTPases; involved in cell signal transduction
NAD <sup>+</sup>	= Nicotinamide adenine dinucleotide	RASSF1A	= Ras association domain-containing protein 1, encoded by the RASSF1 gene
NADP <sup>+</sup>	= Nicotinamide adenine dinucleotide phosphate	RB	= Retinoblastoma protein, also known as pRb; a tumour suppressor protein encoded by the RB1 gene
NADPH	= The reduced form of NADP <sup>+</sup>		
NAMPT	= Nicotinamide phosphoribosyltransferase		
NCI	= National Cancer Institute		
NCI-H727	= A human pulmonary carcinoid cell line		
NF-κB	= Nuclear factor-κB		
NGFR	= Nerve growth factor receptor		
NIH	= National Institutes of Health		
NM	= Nicotinamide		
NMNAT	= Nicotinamide mononucleotide adenylyltransferase		
NMN	= Nicotinamide mononucleotide		
NMNAT	= Nicotinamide mononucleotide adenylyltransferase		

RECIST	= Response evaluation criteria in solid tumours, adopted by, e.g., EORTC, NCI of the USA, and NCI of Canada
RelA	= The <i>RELA</i> gene codes for the transcription factor p65, part of the NF- $\kappa$ B complex
RET	= Receptor tyrosine kinase, a proto-oncogene
ROS	= Reactive oxygen species
RUNX3	= Runt-related transcription factor 3
SCLC	= Small cell lung carcinoma
SAPK	= Stress-activated protein kinase, also known as MAPK9
siRNA	= Small interfering RNA, also called short interfering RNA or silencing, RNA
SMAD	= A class of proteins that modulate the TGF $\beta$ signalling
SMAD2	= One of the receptor-regulated SMADs
SMAD4	= The common-mediator SMAD
SMAD7	= One of the inhibitory SMADs
SOM230	= A pan-somatostatin receptor analogue
SSTRs	= Somatostatin receptors, consisting of 5 subtypes, SSTR1-SSTR5
STAT	= Signal transducers and activator of transcription protein; also called signal, transduction and transcription protein
STS	= A human ileal (midgut) carcinoid cell line
TGF $\alpha$	= Transforming growth factor $\alpha$
TGF $\beta$	= Transforming growth factor $\beta$
TGF $\beta$ BR	= Transforming growth factor $\beta$ receptor; a serine/threonine kinase receptor
TKI	= Tyrosine kinase inhibitor
TNF	= Tumour necrosis factor
TRAIL	= TNF-related apoptosis-inducing ligand, also designated CD253; the natural ligand of TRAIL receptors
TRAILR	= TRAIL receptors
TRAILR1	= TRAIL receptor 1, also called DR4; one of the death receptors
TRAILR2	= TRAIL receptor 2, also called DR5; one of the death receptors
TT	= A human medullary thyroid carcinoma cell line
Tyr	= Tyrosine
UPP	= Ubiquitin-proteasome pathway
VEGF	= Vascular endothelial growth factor; exists in 5 subtypes: VEGFA-VEGFE
VEGFR	= Vascular endothelial growth factor receptor
VEGFR1	= Vascular endothelial growth factor receptor 1, also called FLT1
VEGFR2	= Vascular endothelial growth factor receptor 2, also called KDR/FLK1
VEGFR3	= Vascular endothelial growth factor receptor 3, also called FLT4
VPA	= Valproic acid
WAF1	= Cyclin-dependent kinase inhibitor 1A, also known as p21, CIP1 or CDKN1A, a cell cycle inhibitor
ZM336372	= A RAF1 activator

## REFERENCES

- [1] Ahlman, H.; Olausson, M., Surgery of liver metastases in neuroendocrine tumours. In *Gastrointestinal Oncology: A Critical Multidisciplinary Team Approach*. Blackwell: USA, **2008**; pp 632-638.
- [2] Olausson, M.; Friman, S.; Herlenius, G.; Cahlin, C.; Nilsson, O.; Jansson, S.; Wängberg, B.; Ahlman, H. Orthotopic liver or multivisceral transplantation as treatment of metastatic neuroendocrine tumors. *Liver Transpl.*, **2007**, *13*(3), 327-333.
- [3] Patel, Y. C.; Greenwood, M. T.; Panetta, R.; Demchyshyn, L.; Niznik, H.; Srikant, C. B. The somatostatin receptor family. *Life Sci.*, **1995**, *57*(13), 1249-1265.
- [4] Sharma, K.; Patel, Y. C.; Srikant, C. B. C-terminal region of human somatostatin receptor 5 is required for induction of Rb and G1 cell cycle arrest. *Mol. Endocrinol.*, **1999**, *13*(1), 82-90.
- [5] Pages, P.; Benali, N.; Saint-Laurent, N.; Esteve, J. P.; Schally, A. V.; Tkaczuk, J.; Vaysse, N.; Susini, C.; Buscail, L. sst2 somatostatin receptor mediates cell cycle arrest and induction of p27(Kip1). Evidence for the role of SHP-1. *J. Biol. Chem.*, **1999**, *274*(21), 15186-15193.
- [6] Kölby, L.; Wängberg, B.; Ahlman, H.; Tisel, L. E.; Fjälling, M.; Forssell-Aronsson, E.; Nilsson, O. Somatostatin receptor subtypes, octreotide scintigraphy, and clinical response to octreotide treatment in patients with neuroendocrine tumors. *World J. Surg.*, **1998**, *22*(7), 679-683.
- [7] Weckbecker, G.; Lewis, I.; Albert, R.; Schmid, H. A.; Hoyer, D.; Bruns, C. Opportunities in somatostatin research: biological, chemical and therapeutic aspects. *Nat. Rev. Drug Discov.*, **2003**, *2*(12), 999-1017.
- [8] Boerlin, V.; van der Hoek, J.; Beglinger, C.; Poon, K. W.; Hartmann, S.; Dutreix, C.; Kovarik, J. M.; Bruns, C.; Weckbecker, G.; Lewis, I.; Schnieper, P.; Hofland, L. J.; Lamberts, S. W. New insights on SOM230, a universal somatostatin receptor ligand. *J. Endocrinol. Invest.*, **2003**, *26*(8 Suppl), 14-16.
- [9] Rocheville, M.; Lange, D. C.; Kumar, U.; Patel, S. C.; Patel, R. C.; Patel, Y. C. Receptors for dopamine and somatostatin: formation of hetero-oligomers with enhanced functional activity. *Science*, **2000**, *288*(5463), 154-157.
- [10] Ahlman, H.; Åhlund, L.; Dahlström, A.; Martner, J.; Stenqvist, O.; Tylene, U. SMS 201-995 and provocation tests in preparation of patients with carcinoids for surgery or hepatic arterial embolization. *Anesth. Analg.*, **1988**, *67*(12), 1142-1148.
- [11] Rinke, A.; Müller, H. H.; Schade-Brittinger, C.; Klose, K. J.; Barth, P.; Wied, M.; Mayer, C.; Aminossadati, B.; Pape, U. F.; Blaker, M.; Harder, J.; Arnold, C.; Gress, T.; Arnold, R. Placebo-controlled, double-blind, prospective, randomized study on the effect of octreotide LAR in the control of tumor growth in patients with metastatic neuroendocrine midgut tumors: a report from the PROMID Study Group. *J. Clin. Oncol.*, **2009**, *27*(28), 4656-4663.
- [12] Kwekkeboom, D. J.; de Herder, W. W.; Kam, B. L.; van Eijck, C. H.; van Essen, M.; Kooij, P. P.; Feelders, R. A.; van Aken, M. O.; Krenning, E. P. Treatment with the radiolabeled somatostatin analog [177 Lu-DOTA 0.Tyr3]octreotate: toxicity, efficacy, and survival. *J. Clin. Oncol.*, **2008**, *26*(13), 2124-2130.
- [13] Schally, A. V. New approaches to the therapy of various tumors based on peptide analogues. *Horm. Metab. Res.*, **2008**, *40*(5), 315-322.
- [14] Chaudhry, A.; Funa, K.; Oberg, K. Expression of growth factor peptides and their receptors in neuroendocrine tumors of the digestive system. *Acta. Oncol.*, **1993**, *32*(2), 107-114.
- [15] La Rosa, S.; Uccella, S.; Finzi, G.; Albarello, L.; Sessa, F.; Capella, C. Localization of vascular endothelial growth factor and its receptors in digestive endocrine tumors: correlation with microvessel density and clinicopathologic features. *Hum. Pathol.*, **2003**, *34*(1), 18-27.
- [16] Pavel, M. E., Novel treatment. In *Gastrointestinal Oncology: A Critical Multidisciplinary Team Approach*. Blackwell: USA, **2008**; pp. 720-725.
- [17] Papouchado, B.; Erickson, L. A.; Rohlinger, A. L.; Hobday, T. J.; Erlichman, C.; Ames, M. M.; Lloyd, R. V. Epidermal growth factor receptor and activated epidermal growth factor receptor expression in gastrointestinal carcinoids and pancreatic endocrine carcinomas. *Mod. Pathol.*, **2005**, *18*(10), 1329-1335.

- [18] Hobday, T. J.; Holen, K.; Donehower, R.; Camoriano, J.; Kim, G.; Picus, J.; Philip, P.; Lloyd, R.; Mahoney, M.; Erlichman, C. A phase II trial of gefitinib in patients (pts) with progressive metastatic neuroendocrine tumors (NET): A Phase II Consortium (P2C) study. *J. Clin. Oncol. (Meeting Abstracts)*, **2006**, *24*(18 suppl), 4043-.
- [19] Bourhis, J.; Rivera, F.; Mesia, R.; Awada, A.; Geoffrois, L.; Borel, C.; Humblet, Y.; Lopez-Pousa, A.; Hitt, R.; Vega Villegas, M. E.; Duck, L.; Rosine, D.; Amellal, N.; Schueler, A.; Harstrick, A. Phase I/II study of cetuximab in combination with cisplatin or carboplatin and fluorouracil in patients with recurrent or metastatic squamous cell carcinoma of the head and neck. *J. Clin. Oncol.*, **2006**, *24*(18), 2866-2872.
- [20] Huether, A.; Hopfner, M.; Baradari, V.; Schuppan, D.; Scherubl, H. EGFR blockade by cetuximab alone or as combination therapy for growth control of hepatocellular cancer. *Biochem. Pharmacol.*, **2005**, *70*(11), 1568-78.
- [21] Duax, W. L.; Huether, R.; Pletnev, V. Z.; Langs, D.; Addlagatta, A.; Connare, S.; Habegger, L.; Gill, J. Rational genomics I: antisense open reading frames and codon bias in short-chain oxidoreductase enzymes and the evolution of the genetic code. *Proteins*, **2005**, *61*(4), 900-906.
- [22] Ciardiello, F.; Troiani, T.; Bianco, R.; Oritura, M.; Morgillo, F.; Martinelli, E.; Morelli, M. P.; Cascone, T.; Tortora, G. Interaction between the epidermal growth factor receptor (EGFR) and the vascular endothelial growth factor (VEGF) pathways: a rational approach for multi-target anticancer therapy. *Ann. Oncol.*, **2006**, *17* (Suppl 7), vii109-14.
- [23] Kulke, M. H.; Lenz, H. J.; Meropol, N. J.; Posey, J.; Ryan, D. P.; Picus, J.; Bergsland, E.; Stuart, K.; Tye, L.; Huang, X.; Li, J. Z.; Baum, C. M.; Fuchs, C. S. Activity of sunitinib in patients with advanced neuroendocrine tumors. *J. Clin. Oncol.*, **2008**, *26*(20), 3403-3410.
- [24] Gross, D. J.; Munter, G.; Bitan, M.; Siegal, T.; Gabizon, A.; Weitzen, R.; Merimsky, O.; Ackerstein, A.; Salmon, A.; Sella, A.; Slavin, S. The role of imatinib mesylate (Gleevec) for treatment of patients with malignant endocrine tumors positive for c-kit or PDGF-R. *Endocr. Relat. Cancer*, **2006**, *13*(2), 535-540.
- [25] Yao, J. C.; Zhang, J. X.; Rashid, A.; Yeung, S. C.; Szklaruk, J.; Hess, K.; Xie, K.; Ellis, L.; Abbruzzese, J. L.; Ajani, J. A. Clinical and *in vitro* studies of imatinib in advanced carcinoid tumors. *Clin. Cancer Res.*, **2007**, *13*(1), 234-40.
- [26] McAuliffe, J. C.; Lazar, A. J.; Yang, D.; Steinert, D. M.; Qiao, W.; Thall, P. F.; Raymond, A. K.; Benjamin, R. S.; Trent, J. C. Association of intratumoral vascular endothelial growth factor expression and clinical outcome for patients with gastrointestinal stromal tumors treated with imatinib mesylate. *Clin. Cancer Res.*, **2007**, *13*(22 Pt 1), 6727-6734.
- [27] Nilsson, B.; Nilsson, O.; Ahlman, H. Treatment of gastrointestinal stromal tumours: imatinib, sunitinib -- and then? *Expert Opin. Investig. Drugs*, **2009**, *18*(4), 457-468.
- [28] Gazdar, A. F. Activating and resistance mutations of EGFR in non-small-cell lung cancer: role in clinical response to EGFR tyrosine kinase inhibitors. *Oncogene*, **2009**, *28*(Suppl 1), S24-31.
- [29] Wang, L.; Ignat, A.; Axiotis, C. A. Differential expression of the PTEN tumor suppressor protein in fetal and adult neuroendocrine tissues and tumors: progressive loss of PTEN expression in poorly differentiated neuroendocrine neoplasms. *Appl. Immunohistochem. Mol. Morphol.*, **2002**, *10*(2), 139-146.
- [30] Perren, A.; Komminoth, P.; Saremaslani, P.; Matter, C.; Feurer, S.; Lees, J. A.; Heitz, P. U.; Eng, C. Mutation and expression analyses reveal differential subcellular compartmentalization of PTEN in endocrine pancreatic tumors compared to normal islet cells. *Am. J. Pathol.*, **2000**, *157*(4), 1097-1103.
- [31] Zitzmann, K.; De Toni, E. N.; Brand, S.; Goke, B.; Meinecke, J.; Spottl, G.; Meyer, H. H.; Auernhammer, C. J. The novel mTOR inhibitor RAD001 (everolimus) induces antiproliferative effects in human pancreatic neuroendocrine tumor cells. *Neuroendocrinology*, **2007**, *85*(1), 54-60.
- [32] Yao, J. C.; Phan, A. T.; Chang, D. Z.; Wolff, R. A.; Hess, K.; Gupta, S.; Jacobs, C.; Mares, J. E.; Landgraf, A. N.; Rashid, A.; Meric-Bernstam, F. Efficacy of RAD001 (everolimus) and octreotide LAR in advanced low- to intermediate-grade neuroendocrine tumors: results of a phase II study. *J. Clin. Oncol.*, **2008**, *26*(26), 4311-8.
- [33] Yao, J. C.; Lombard-Bohas, C.; Baudin, E.; Kvols, L. K.; Rougier, P.; Ruzsniwski, P.; Hoosen, S.; St Peter, J.; Haas, T.; Lebowitz, D.; Van Cutsem, E.; Kulke, M. H.; Hobday, T. J.; O'Dorisio, T. M.; Shah, M. H.; Cadiot, G.; Luppi, G.; Posey, J. A.; Wiedenmann, B. Daily oral everolimus activity in patients with metastatic pancreatic neuroendocrine tumors after failure of cytotoxic chemotherapy: a phase II trial. *J. Clin. Oncol.*, **2010**, *28*(1), 69-76.
- [34] Terris, B.; Scoazec, J. Y.; Rubbia, L.; Bregeud, L.; Pepper, M. S.; Ruzsniwski, P.; Belghiti, J.; Flejou, J.; Degott, C. Expression of vascular endothelial growth factor in digestive neuroendocrine tumours. *Histopathology*, **1998**, *32*(2), 133-138.
- [35] Pavel, M. E.; Hassler, G.; Baum, U.; Hahn, E. G.; Lohmann, T.; Schuppan, D. Circulating levels of angiogenic cytokines can predict tumour progression and prognosis in neuroendocrine carcinomas. *Clin. Endocrinol. (Oxf)*, **2005**, *62*(4), 434-443.
- [36] Zhang, J.; Jia, Z.; Li, Q.; Wang, L.; Rashid, A.; Zhu, Z.; Evans, D. B.; Vauthey, J. N.; Xie, K.; Yao, J. C. Elevated expression of vascular endothelial growth factor correlates with increased angiogenesis and decreased progression-free survival among patients with low-grade neuroendocrine tumors. *Cancer*, **2007**, *109*(8), 1478-86.
- [37] Hurwitz, H.; Fehrenbacher, L.; Novotny, W.; Cartwright, T.; Hainsworth, J.; Heim, W.; Berlin, J.; Baron, A.; Griffing, S.; Holmgren, E.; Ferrara, N.; Fyfe, G.; Rogers, B.; Ross, R.; Kabbinavar, F. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *N. Engl. J. Med.*, **2004**, *350*(23), 2335-42.
- [38] Yao, J. C.; Phan, A.; Hoff, P. M.; Chen, H. X.; Charnsangavej, C.; Yeung, S. C.; Hess, K.; Ng, C.; Abbruzzese, J. L.; Ajani, J. A. Targeting vascular endothelial growth factor in advanced carcinoid tumor: a random assignment phase II study of depot octreotide with bevacizumab and pegylated interferon alpha-2b. *J. Clin. Oncol.*, **2008**, *26*(8), 1316-23.
- [39] Cicek, M.; Iwaniec, U. T.; Goblirsch, M. J.; Vrabell, A.; Ruan, M.; Clohisy, D. R.; Turner, R. R.; Oursler, M. J. 2-Methoxyestradiol suppresses osteolytic breast cancer tumor progression *in vivo*. *Cancer Res.*, **2007**, *67*(21), 10106-10111.
- [40] Dahut, W. L.; Lakhani, N. J.; Gulley, J. L.; Arlen, P. M.; Kohn, E. C.; Kotz, H.; McNally, D.; Parr, A.; Nguyen, D.; Yang, S. X.; Steinberg, S. M.; Venitz, J.; Sparreboom, A.; Figg, W. D. Phase I clinical trial of oral 2-methoxyestradiol, an antiangiogenic and apoptotic agent, in patients with solid tumors. *Cancer Biol. Ther.*, **2006**, *5*(1), 22-27.
- [41] Rosen, L. S.; Kurzrock, R.; Mulay, M.; Van Vugt, A.; Purdom, M.; Ng, C.; Silverman, J.; Koutsoukos, A.; Sun, Y. N.; Bass, M. B.; Xu, R. Y.; Polverino, A.; Wiezorek, J. S.; Chang, D. D.; Benjamin, R.; Herbst, R. S. Safety, pharmacokinetics, and efficacy of AMG 706, an oral multikinase inhibitor, in patients with advanced solid tumors. *J. Clin. Oncol.*, **2007**, *25*(17), 2369-2376.
- [42] Kumar, R.; Knick, V. B.; Rudolph, S. K.; Johnson, J. H.; Crosby, R. M.; Crouthamel, M. C.; Hopper, T. M.; Miller, C. G.; Harrington, L. E.; Onori, J. A.; Mullin, R. J.; Gilmer, T. M.; Truesdale, A. T.; Epperly, A. H.; Bolor, A.; Stafford, J. A.; Luttrell, D. K.; Cheung, M. Pharmacokinetic-pharmacodynamic correlation from mouse to human with pazopanib, a multikinase angiogenesis inhibitor with potent antitumor and antiangiogenic activity. *Mol. Cancer Ther.*, **2007**, *6*(7), 2012-2021.
- [43] Fischer, C.; Jonckx, B.; Mazzone, M.; Zacchigna, S.; Loges, S.; Pattarini, L.; Chorianopoulos, E.; Liesenborghs, L.; Koch, M.; De Mol, M.; Autiero, M.; Wynn, S.; Plaisance, S.; Moons, L.; van Rooijen, N.; Giacca, M.; Stassen, J. M.; Dewerchin, M.; Collen, D.; Carmeliet, P. Anti-PIGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. *Cell*, **2007**, *131*(3), 463-475.
- [44] Kulke, M. H.; Stuart, K.; Enzinger, P. C.; Ryan, D. P.; Clark, J. W.; Muzikansky, A.; Vincitore, M.; Michelini, A.; Fuchs, C. S. Phase II study of temozolomide and thalidomide in patients with metastatic neuroendocrine tumors. *J. Clin. Oncol.*, **2006**, *24*(3), 401-406.
- [45] Kulke, M. H.; Bergsland, E. K.; Ryan, D. P.; Enzinger, P. C.; Lynch, T. J.; Zhu, A. X.; Meyerhardt, J. A.; Heymach, J. V.; Fogler, W. E.; Sidor, C.; Michelini, A.; Kinsella, K.; Venook, A. P.; Fuchs, C. S. Phase II study of recombinant human endostatin in

- patients with advanced neuroendocrine tumors. *J. Clin. Oncol.*, **2006**, *24*(22), 3555-3561.
- [46] Modlin, I. M.; Moss, S. F.; Chung, D. C.; Jensen, R. T.; Snyderwine, E. Priorities for improving the management of gastroenteropancreatic neuroendocrine tumors. *J. Natl. Cancer Inst.*, **2008**, *100*(18), 1282-1289.
- [47] Evers, B. M.; Townsend, C. M., Jr.; Upp, J. R.; Allen, E.; Hurlbut, S. C.; Kim, S. W.; Rajaraman, S.; Singh, P.; Reubi, J. C.; Thompson, J. C. Establishment and characterization of a human carcinoid in nude mice and effect of various agents on tumor growth. *Gastroenterology*, **1991**, *101*(2), 303-311.
- [48] Pfragner, R.; Wirnsberger, G.; Niederle, B.; Behmel, A.; Rinner, I.; Mandl, A.; Wavrina, F.; Luo, J. S.; Adamiker, D.; Hoegner, H.; Ingolic, E.; Schauenstein, K. Establishment of a continuous cell line from a human carcinoid of the small intestine (KRJ-I): characterization and effects of 5-azacytidine on proliferation. *Int. J. Oncol.*, **1996**, *8*, 513-520.
- [49] Kölby, L.; Bernhardt, P.; Ahlman, H.; Wängberg, B.; Johanson, V.; Wigander, A.; Forssell-Aronsson, E.; Karlsson, S.; Ahren, B.; Stenman, G.; Nilsson, O. A transplantable human carcinoid as model for somatostatin receptor-mediated and amine transporter-mediated radionuclide uptake. *Am. J. Pathol.*, **2001**, *158*(2), 745-755.
- [50] Van Buren, G.; Rashid, A.; Yang, A. D.; Abdalla, E. K.; Gray, M. J.; Liu, W.; Somcio, R.; Fan, F.; Camp, E. R.; Yao, J. C.; Ellis, L. M. The development and characterization of a human midgut carcinoid cell line. *Clin. Cancer Res.*, **2007**, *13*(16), 4704-4712.
- [51] Pfragner, R.; Behmel, A.; Hoger, H.; Beham, A.; Ingolic, E.; Stelzer, I.; Svejda, B.; Moser, V. A.; Obenaus, A. C.; Siegl, V.; Haas, O.; Niederle, B. Establishment and characterization of three novel cell lines - P-ST5, L-ST5, H-ST5 - derived from a human metastatic midgut carcinoid. *Anticancer Res.*, **2009**, *29*(6), 1951-1961.
- [52] Nilsson, O.; Kolby, L.; Bernhardt, P.; Forssell-Aronsson, E.; Johanson, V.; Ahlman, H. GOT1 xenografted to nude mice: a unique model for *in vivo* studies on SSTR-mediated radiation therapy of carcinoid tumors. *Ann. N Y Acad. Sci.*, **2004**, *1014*, 275-279.
- [53] Siddique, Z. L.; Drozdov, I.; Floch, J.; Gustafsson, B. I.; Stunes, K.; Pfragner, R.; Kidd, M.; Modlin, I. M. KRJ-I and BON cell lines: defining an appropriate enterochromaffin cell neuroendocrine tumor model. *Neuroendocrinology*, **2009**, *89*(4), 458-470.
- [54] Sethi, G.; Sung, B.; Aggarwal, B. B. Nuclear Factor- $\kappa$ B Activation: From Bench to Bedside. *Exp. Biol. Med.*, **2008**, *233*(1), 21-31.
- [55] Namba, H.; Saenko, V.; Yamashita, S. Nuclear factor- $\kappa$ B in thyroid carcinogenesis and progression: a novel therapeutic target for advanced thyroid cancer. *Arq. Bras. Endocrinol. Metabol.*, **2007**, *51*(5), 843-851.
- [56] Suh, J.; Payvandi, F.; Edelstein, L. C.; Amenta, P. S.; Zong, W. X.; Gelinas, C.; Rabson, A. B. Mechanisms of constitutive NF- $\kappa$ B activation in human prostate cancer cells. *Prostate*, **2002**, *52*(3), 183-200.
- [57] Kojima, M.; Morisaki, T.; Sasaki, N.; Nakano, K.; Mibu, R.; Tanaka, M.; Katano, M. Increased nuclear factor- $\kappa$ B activation in human colorectal carcinoma and its correlation with tumor progression. *Anticancer Res.*, **2004**, *24*(2B), 675-681.
- [58] Wang, W.; Abbuzzese, J. L.; Evans, D. B.; Larry, L.; Cleary, K. R.; Chiao, P. J. The nuclear factor- $\kappa$ B RelA transcription factor is constitutively activated in human pancreatic adenocarcinoma cells. *Clin. Cancer Res.*, **1999**, *5*(1), 119-127.
- [59] Levidou, G.; Saetta, A. A.; Korkolopoulou, P.; Papanastasiou, P.; Gioti, K.; Pavlopoulos, P.; Diamantopoulou, K.; Thomas-Tsagli, E.; Xiromeritis, K.; Patsouris, E. Clinical significance of nuclear factor (NF)- $\kappa$ B levels in urothelial carcinoma of the urinary bladder. *Virchows Arch.*, **2008**, *452*(3), 295-304.
- [60] Sweeney, C.; Li, L.; Shanmugam, R.; Bhat-Nakshatri, P.; Jayaprakasan, V.; Baldrige, L. A.; Gardner, T.; Smith, M.; Nakshatri, H.; Cheng, L. Nuclear factor- $\kappa$ B is constitutively activated in prostate cancer *in vitro* and is overexpressed in prostatic intraepithelial neoplasia and adenocarcinoma of the prostate. *Clin. Cancer Res.*, **2004**, *10*(16), 5501-7.
- [61] Krappmann, D.; Emmerich, F.; Kordes, U.; Scharshmidt, E.; Dorken, B.; Scheiderei, C. Molecular mechanisms of constitutive NF- $\kappa$ B/Rel activation in Hodgkin/Reed-Sternberg cells. *Oncogene*, **1999**, *18*(4), 943-953.
- [62] Karin, M.; Yamamoto, Y.; Wang, Q. M. The IKK NF- $\kappa$ B system: a treasure trove for drug development. *Nat. Rev. Drug Discov.*, **2004**, *3*(1), 17-26.
- [63] Olsen, L. S.; Hjarnaa, P. J.; Latini, S.; Holm, P. K.; Larsson, R.; Bramm, E.; Binderup, L.; Madsen, M. W. Anticancer agent CHS 828 suppresses nuclear factor- $\kappa$ B activity in cancer cells through downregulation of IKK activity. *Int. J. Cancer*, **2004**, *111*(2), 198-205.
- [64] Ludwig, L.; Kessler, H.; Wagner, M.; Hoang-Vu, C.; Dralle, H.; Adler, G.; Bohm, B. O.; Schmid, R. M. Nuclear factor- $\kappa$ B is constitutively active in C-cell carcinoma and required for RET-induced transformation. *Cancer Res.*, **2001**, *61*(11), 4526-4535.
- [65] Svensson, A.; Backman, U.; Jonsson, E.; Larsson, R.; Christofferson, R. CHS 828 inhibits neuroblastoma growth in mice alone and in combination with antiangiogenic drugs. *Pediatr. Res.*, **2002**, *51*(5), 607-11.
- [66] Johanson, V.; Arvidsson, Y.; Kölby, L.; Bernhardt, P.; Swärd, C.; Nilsson, O.; Ahlman, H. Antitumoural effects of the pyridyl cyanoguanidine CHS 828 on three different types of neuroendocrine tumours xenografted to nude mice. *Neuroendocrinology*, **2005**, *82*(3-4), 171-176.
- [67] Olesen, U. H.; Christensen, M. K.; Bjorkling, F.; Jaattela, M.; Jensen, P. B.; Sehested, M.; Nielsen, S. J. Anticancer agent CHS-828 inhibits cellular synthesis of NAD. *Biochem. Biophys. Res. Commun.*, **2008**, *367*(4), 799-804.
- [68] Hassa, P. O.; Haenni, S. S.; Elser, M.; Hottiger, M. O. Nuclear ADP-ribosylation reactions in mammalian cells: where are we today and where are we going? *Microbiol. Mol. Biol. Rev.*, **2006**, *70*(3), 789-829.
- [69] Schreiber, V.; Dantzer, F.; Ame, J. C.; de Murcia, G. Poly(ADP-ribose): novel functions for an old molecule. *Nat. Rev. Mol. Cell Biol.*, **2006**, *7*(7), 517-528.
- [70] Khan, J. A.; Forouhar, F.; Tao, X.; Tong, L. Nicotinamide adenine dinucleotide metabolism as an attractive target for drug discovery. *Expert Opin. Ther. Targets*, **2007**, *11*(5), 695-705.
- [71] Berger, N. A. Poly(ADP-ribose) in the cellular response to DNA damage. *Radiat. Res.*, **1985**, *101*(1), 4-15.
- [72] Hasmann, M.; Schemainda, I. FK866, a highly specific noncompetitive inhibitor of nicotinamide phosphoribosyltransferase, represents a novel mechanism for induction of tumor cell apoptosis. *Cancer Res.*, **2003**, *63*(21), 7436-7442.
- [73] Watson, M.; Roulston, A.; Belec, L.; Billot, X.; Marcellus, R.; Bedard, D.; Bernier, C.; Branchaud, S.; Chan, H.; Dairi, K.; Gilbert, K.; Goulet, D.; Gratton, M. O.; Isakau, H.; Jang, A.; Khadir, A.; Koch, E.; Lavoie, M.; Lawless, M.; Nguyen, M.; Paquette, D.; Turcotte, E.; Berger, A.; Mitchell, M.; Shore, G. C.; Beuparlant, P. The small molecule GMX1778 is a potent inhibitor of NAD<sup>+</sup> biosynthesis: strategy for enhanced therapy in nicotinic acid phosphoribosyltransferase 1-deficient tumors. *Mol. Cell Biol.*, **2009**, *29*(21), 5872-5888.
- [74] von Heideman, A.; Berglund, A.; Larsson, R.; Nygren, P. Safety and efficacy of NAD depleting cancer drugs: results of a phase I clinical trial of CHS 828 and overview of published data. *Cancer Chemother. Pharmacol.*, **2009**, *65*(6), 1165-1172.
- [75] Roccaro, A. M.; Hideshima, T.; Richardson, P. G.; Russo, D.; Ribatti, D.; Vacca, A.; Dammacco, F.; Anderson, K. C. Bortezomib as an antitumor agent. *Curr. Pharm. Biotechnol.*, **2006**, *7*(6), 441-448.
- [76] Larsson, D. E.; Lovborg, H.; Rickardson, L.; Larsson, R.; Öberg, K.; Granberg, D. Identification and evaluation of potential anti-cancer drugs on human neuroendocrine tumor cell lines. *Anticancer Res.*, **2006**, *26*(6B), 4125-4129.
- [77] Shah, M. H.; Young, D.; Kindler, H. L.; Webb, I.; Kleiber, B.; Wright, J.; Grever, M. Phase II study of the proteasome inhibitor bortezomib (PS-341) in patients with metastatic neuroendocrine tumors. *Clin. Cancer Res.*, **2004**, *10*(18 Pt 1), 6111-6118.
- [78] Baradari, V.; Hopfner, M.; Huether, A.; Schuppan, D.; Scherubl, H. Histone deacetylase inhibitor MS-275 alone or combined with bortezomib or sorafenib exhibits strong antiproliferative action in human cholangiocarcinoma cells. *World J. Gastroenterol.*, **2007**, *13*(33), 4458-4466.

- [79] Harris, A. L. Chemotherapy for the carcinoid syndrome. *Cancer Chemother. Pharmacol.*, **1981**, *5*(3), 133-138.
- [80] Moertel, C. G. Treatment of the carcinoid tumor and the malignant carcinoid syndrome. *J. Clin. Oncol.*, **1983**, *1*(11), 727-740.
- [81] Chakravarthy, A.; Abrams, R. A. Radiation therapy in the management of patients with malignant carcinoid tumors. *Cancer*, **1995**, *75*(6), 1386-1390.
- [82] Kulke, M. H.; Mayer, R. J. Carcinoid tumors. *N. Engl. J. Med.*, **1999**, *340*(11), 858-868.
- [83] Kimmig, B. N. Radiotherapy for gastroenteropancreatic neuroendocrine tumors. *Ann. N Y Acad. Sci.*, **1994**, *733*, 488-495.
- [84] Horton, K. M.; Kamel, I.; Hofmann, L.; Fishman, E. K. Carcinoid tumors of the small bowel: a multitechnique imaging approach. *Am. J. Roentgenol.*, **2004**, *182*(3), 559-67.
- [85] Samlowski, W. E.; Eyre, H. J.; Sause, W. T. Evaluation of the response of unresectable carcinoid tumors to radiotherapy. *Int. J. Radiat. Oncol. Biol. Phys.*, **1986**, *12*(3), 301-305.
- [86] Abrams, R. A.; King, D.; Wilson, J. F. Objective response of malignant carcinoid to radiation therapy. *Int. J. Radiat. Oncol. Biol. Phys.*, **1987**, *13*(6), 869-873.
- [87] Keane, T. J.; Rider, W. D.; Harwood, A. R.; Thomas, G. M.; Cummings, B. J. Whole abdominal radiation in the management of metastatic gastrointestinal carcinoid tumor. *Int. J. Radiat. Oncol. Biol. Phys.*, **1981**, *7*(11), 1519-1521.
- [88] Schupak, K. D.; Wallner, K. E. The role of radiation therapy in the treatment of locally unresectable or metastatic carcinoid tumors. *Int. J. Radiat. Oncol. Biol. Phys.*, **1991**, *20*(3), 489-495.
- [89] van Essen, M.; Krenning, E. P.; Kam, B. L.; de Jong, M.; Valkema, R.; Kwekkeboom, D. J. Peptide-receptor radionuclide therapy for endocrine tumors. *Nat. Rev. Endocrinol.*, **2009**, *5*(7), 382-393.
- [90] Kölby, L.; Bernhardt, P.; Johanson, V.; Schmitt, A.; Ahlman, H.; Forssell-Aronsson, E.; Mäcke, H.; Nilsson, O. Successful receptor-mediated radiation therapy of xenografted human midgut carcinoid tumour. *Br. J. Cancer*, **2005**, *93*(10), 1144-1151.
- [91] Oddstig, J.; Bernhardt, P.; Nilsson, O.; Ahlman, H.; Forssell-Aronsson, E. Radiation-induced up-regulation of somatostatin receptor expression in small cell lung cancer *in vitro*. *Nucl. Med. Biol.*, **2006**, *33*(7), 841-846.
- [92] Bernhardt, P.; Oddstig, J.; Kölby, L.; Nilsson, O.; Ahlman, H.; Forssell-Aronsson, E. Effects of treatment with (177)Lu-DOTA-Tyr(3)-octreotate on uptake of subsequent injection in carcinoid-bearing nude mice. *Cancer Biother. Radiopharm.*, **2007**, *22*(5), 644-653.
- [93] Verheij, M.; Bartelink, H. Radiation-induced apoptosis. *Cell Tissue Res.*, **2000**, *301*(1), 133-142.
- [94] Shinomiya, N. New concepts in radiation-induced apoptosis: "premitotic apoptosis" and "postmitotic apoptosis". *J. Cell Mol. Med.*, **2001**, *5*(3), 240-253.
- [95] Hamada, N.; Matsumoto, H.; Hara, T.; Kobayashi, Y. Intercellular and intracellular signaling pathways mediating ionizing radiation-induced bystander effects. *J. Radiat. Res. (Tokyo)*, **2007**, *48*(2), 87-95.
- [96] Zhou, H.; Ivanov, V. N.; Gillespie, J.; Geard, C. R.; Amundson, S. A.; Brenner, D. J.; Yu, Z.; Lieberman, H. B.; Hei, T. K. Mechanism of radiation-induced bystander effect: role of the cyclooxygenase-2 signaling pathway. *Proc. Natl. Acad. Sci. USA*, **2005**, *102*(41), 14641-14646.
- [97] Azzam, E. I.; De Toledo, S. M.; Spitz, D. R.; Little, J. B. Oxidative metabolism modulates signal transduction and micronucleus formation in bystander cells from alpha-particle-irradiated normal human fibroblast cultures. *Cancer Res.*, **2002**, *62*(19), 5436-5442.
- [98] Zhou, H.; Suzuki, M.; Randers-Pehrson, G.; Vannais, D.; Chen, G.; Trosko, J. E.; Waldren, C. A.; Hei, T. K. Radiation risk to low fluences of alpha particles may be greater than we thought. *Proc. Natl. Acad. Sci. USA*, **2001**, *98*(25), 14410-14415.
- [99] Cadden, I.; Johnston, B. T.; Turner, G.; McCance, D.; Ardill, J.; McGinty, A. An evaluation of cyclooxygenase-2 as a prognostic biomarker in mid-gut carcinoid tumours. *Neuroendocrinology*, **2007**, *86*(2), 104-111.
- [100] Schulze-Bergkamen, H.; Weinmann, A.; Moehler, M.; Siebler, J.; Galle, P. R. Novel ways to sensitize gastrointestinal cancer to apoptosis. *Gut*, **2009**, *58*(7), 1010-1024.
- [101] Wang, D. G. Apoptosis in neuroendocrine tumours. *Clin. Endocrinol. (Oxf)*, **1999**, *51*(1), 1-9.
- [102] Wang, D. G.; Johnston, C. F.; Anderson, N.; Sloan, J. M.; Buchanan, K. D. Overexpression of the tumour suppressor gene p53 is not implicated in neuroendocrine tumour carcinogenesis. *J. Pathol.*, **1995**, *175*(4), 397-401.
- [103] Pasquini, L.; Petrucci, E.; Riccioni, R.; Petronelli, A.; Testa, U. Sensitivity and resistance of human cancer cells to TRAIL: mechanisms and therapeutical perspectives. *Cancer Ther.*, **2006**, *4*, 47-72.
- [104] Griffith, T. S.; Stokes, B.; Kucaba, T. A.; Earel, J. K., Jr.; VanOosten, R. L.; Brincks, E. L.; Norian, L. A. TRAIL gene therapy: from preclinical development to clinical application. *Curr. Gene Ther.*, **2009**, *9*(1), 9-19.
- [105] Johnstone, R. W.; Frew, A. J.; Smyth, M. J. The TRAIL apoptotic pathway in cancer onset, progression and therapy. *Nat. Rev. Cancer*, **2008**, *8*(10), 782-798.
- [106] Ashkenazi, A.; Dixit, V. M. Death receptors: signaling and modulation. *Science*, **1998**, *281*(5381), 1305-1308.
- [107] Kim, Y.; Suh, N.; Sporn, M.; Reed, J. C. An inducible pathway for degradation of FLIP protein sensitizes tumor cells to TRAIL-induced apoptosis. *J. Biol. Chem.*, **2002**, *277*(25), 22320-22329.
- [108] Schuchmann, M.; Schulze-Bergkamen, H.; Fleischer, B.; Schattenberg, J. M.; Siebler, J.; Weinmann, A.; Teufel, A.; Worns, M.; Fischer, T.; Strand, S.; Lohse, A. W.; Galle, P. R. Histone deacetylase inhibition by valproic acid down-regulates c-FLIP/CASH and sensitizes hepatoma cells towards CD95- and TRAIL receptor-mediated apoptosis and chemotherapy. *Oncol. Rep.*, **2006**, *15*(1), 227-30.
- [109] Koschny, R.; Walczak, H.; Ganten, T. M. The promise of TRAIL--potential and risks of a novel anticancer therapy. *J. Mol. Med.*, **2007**, *85*(9), 923-935.
- [110] Goke, R.; Goke, A.; Goke, B.; El-Deiry, W. S.; Chen, Y. Pioglitazone inhibits growth of carcinoid cells and promotes TRAIL-induced apoptosis by induction of p21waf1/cip1. *Digestion*, **2001**, *64*(2), 75-80.
- [111] Kuwabara, M.; Takahashi, K.; Inanami, O. Induction of apoptosis through the activation of SAPK/JNK followed by the expression of death receptor Fas in X-irradiated cells. *J. Radiat. Res. (Tokyo)*, **2003**, *44*(3), 203-209.
- [112] Inanami, O.; Takahashi, K.; Kuwabara, M. Attenuation of caspase-3-dependent apoptosis by Trolox post-treatment of X-irradiated MOLT-4 cells. *Int. J. Radiat. Biol.*, **1999**, *75*(2), 155-163.
- [113] McKenna, W. G.; Muschel, R. J.; Gupta, A. K.; Hahn, S. M.; Bernhard, E. J. The RAS signal transduction pathway and its role in radiation sensitivity. *Oncogene*, **2003**, *22*(37), 5866-5875.
- [114] Ling, C. C.; Endlich, B. Radioresistance induced by oncogenic transformation. *Radiat Res.*, **1989**, *120*(2), 267-79.
- [115] Bernhard, E. J.; Kao, G.; Cox, A. D.; Sebti, S. M.; Hamilton, A. D.; Muschel, R. J.; McKenna, W. G. The farnesyltransferase inhibitor FTI-277 radiosensitizes H-ras-transformed rat embryo fibroblasts. *Cancer Res.*, **1996**, *56*(8), 1727-30.
- [116] Miller, A. C.; Kariko, K.; Myers, C. E.; Clark, E. P.; Samid, D. Increased radioresistance of EJras-transformed human osteosarcoma cells and its modulation by lovastatin, an inhibitor of p21ras isoprenylation. *Int. J. Cancer*, **1993**, *53*(2), 302-7.
- [117] Lettini, A. A.; Guidoboni, M.; Fonsatti, E.; Anzalone, L.; Cortini, E.; Maio, M. Epigenetic remodelling of DNA in cancer. *Histol. Histopathol.*, **2007**, *22*(12), 1413-1424.
- [118] Liu, L.; Broadus, R. R.; Yao, J. C.; Xie, S.; White, J. A.; Wu, T. T.; Hamilton, S. R.; Rashid, A. Epigenetic alterations in neuroendocrine tumors: methylation of RAS-association domain family 1, isoform A and p16 genes are associated with metastasis. *Mod. Pathol.*, **2005**, *18*(12), 1632-1640.
- [119] Arnold, C. N.; Sosnowski, A.; Schmitt-Graff, A.; Arnold, R.; Blum, H. E. Analysis of molecular pathways in sporadic neuroendocrine tumors of the gastro-entero-pancreatic system. *Int. J. Cancer*, **2007**, *120*(10), 2157-2164.
- [120] Zhang, H. Y.; Rumilla, K. M.; Jin, L.; Nakamura, N.; Stilling, G. A.; Ruebel, K. H.; Hobday, T. J.; Erlichman, C.; Erickson, L. A.; Lloyd, R. V. Association of DNA methylation and epigenetic inactivation of RASSF1A and beta-catenin with metastasis in small bowel carcinoid tumors. *Endocrine*, **2006**, *30*(3), 299-306.
- [121] Choi, I. S.; Estecio, M. R.; Nagano, Y.; Kim do, H.; White, J. A.; Yao, J. C.; Issa, J. P.; Rashid, A. Hypomethylation of LINE-1 and Alu in well-differentiated neuroendocrine tumors (pancreatic

- endocrine tumors and carcinoid tumors). *Mod. Pathol.*, **2007**, *20*(7), 802-810.
- [122] Bolden, J. E.; Peart, M. J.; Johnstone, R. W. Anticancer activities of histone deacetylase inhibitors. *Nat. Rev. Drug Discov.*, **2006**, *5*(9), 769-784.
- [123] Qi, R.; An, H.; Yu, Y.; Zhang, M.; Liu, S.; Xu, H.; Guo, Z.; Cheng, T.; Cao, X. Notch1 signaling inhibits growth of human hepatocellular carcinoma through induction of cell cycle arrest and apoptosis. *Cancer Res.*, **2003**, *63*(23), 8323-8329.
- [124] Sriuranpong, V.; Borges, M. W.; Ravi, R. K.; Arnold, D. R.; Nelkin, B. D.; Baylin, S. B.; Ball, D. W. Notch signaling induces cell cycle arrest in small cell lung cancer cells. *Cancer Res.*, **2001**, *61*(7), 3200-3205.
- [125] Sippel, R. S.; Carpenter, J. E.; Kunnimalaiyaan, M.; Chen, H. The role of human achaete-scute homolog-1 in medullary thyroid cancer cells. *Surgery*, **2003**, *134*(6), 866-871; discussion 871-3.
- [126] Nakakura, E. K.; Sriuranpong, V. R.; Kunnimalaiyaan, M.; Hsiao, E. C.; Schuebel, K. E.; Borges, M. W.; Jin, N.; Collins, B. J.; Nelkin, B. D.; Chen, H.; Ball, D. W. Regulation of neuroendocrine differentiation in gastrointestinal carcinoid tumor cells by notch signaling. *J. Clin. Endocrinol. Metab.*, **2005**, *90*(7), 4350-4356.
- [127] Roncarati, R.; Sestan, N.; Scheinfeld, M. H.; Berechid, B. E.; Lopez, P. A.; Meucci, O.; McGlade, J. C.; Rakic, P.; D'Adamo, L. The gamma-secretase-generated intracellular domain of beta-amyloid precursor protein binds Numb and inhibits Notch signaling. *Proc. Natl. Acad. Sci. USA*, **2002**, *99*(10), 7102-7107.
- [128] Fischer, D. F.; van Dijk, R.; Sluijs, J. A.; Nair, S. M.; Racchi, M.; Levelt, C. N.; van Leeuwen, F. W.; Hol, E. M. Activation of the Notch pathway in Down syndrome: cross-talk of Notch and APP. *FASEB J.*, **2005**, *19*(11), 1451-1458.
- [129] Arvidsson, Y.; Andersson, E.; Bergström, A.; Andersson, M. K.; Altiparmak, G.; Illerskog, A. C.; Ahlman, H.; Lamazhapova, D.; Nilsson, O. Amyloid precursor-like protein 1 is differentially upregulated in neuroendocrine tumours of the gastrointestinal tract. *Endocr. Relat. Cancer*, **2008**, *15*(2), 569-581.
- [130] Stockhausen, M. T.; Sjölund, J.; Manetopoulos, C.; Axelson, H. Effects of the histone deacetylase inhibitor valproic acid on Notch signalling in human neuroblastoma cells. *Br. J. Cancer*, **2005**, *92*(4), 751-759.
- [131] Greenblatt, D. Y.; Vaccaro, A. M.; Jaskula-Sztul, R.; Ning, L.; Haymart, M.; Kunnimalaiyaan, M.; Chen, H. Valproic acid activates notch-1 signaling and regulates the neuroendocrine phenotype in carcinoid cancer cells. *Oncologist*, **2007**, *12*(8), 942-951.
- [132] Kummur, S.; Gutierrez, M.; Gardner, E. R.; Donovan, E.; Hwang, K.; Chung, E. J.; Lee, M. J.; Maynard, K.; Kalnitskiy, M.; Chen, A.; Melillo, G.; Ryan, Q. C.; Conley, B.; Figg, W. D.; Trepel, J. B.; Zwiebel, J.; Doroshow, J. H.; Murgo, A. J. Phase I trial of MS-275, a histone deacetylase inhibitor, administered weekly in refractory solid tumors and lymphoid malignancies. *Clin. Cancer Res.*, **2007**, *13*(18 Pt 1), 5411-5417.
- [133] Shah, M. H.; Binkley, P.; Chan, K.; Xiao, J.; Arbogast, D.; Collamore, M.; Farra, Y.; Young, D.; Grever, M. Cardiotoxicity of histone deacetylase inhibitor depsipeptide in patients with metastatic neuroendocrine tumors. *Clin. Cancer Res.*, **2006**, *12*(13), 3997-4003.
- [134] Muller, A.; Homey, B.; Soto, H.; Ge, N.; Catron, D.; Buchanan, M. E.; McClanahan, T.; Murphy, E.; Yuan, W.; Wagner, S. N.; Barrera, J. L.; Mohar, A.; Verastegui, E.; Zlotnik, A. Involvement of chemokine receptors in breast cancer metastasis. *Nature*, **2001**, *410*(6824), 50-56.
- [135] Taichman, R. S.; Cooper, C.; Keller, E. T.; Pienta, K. J.; Taichman, N. S.; McCauley, L. K. Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone. *Cancer Res.*, **2002**, *62*(6), 1832-1837.
- [136] Kang, Y.; Siegel, P. M.; Shu, W.; Drobniak, M.; Kakonen, S. M.; Cordon-Cardo, C.; Guise, T. A.; Massague, J. A multigenic program mediating breast cancer metastasis to bone. *Cancer Cell*, **2003**, *3*(6), 537-549.
- [137] Balkwill, F. The significance of cancer cell expression of the chemokine receptor CXCR4. *Semin. Cancer Biol.*, **2004**, *14*(3), 171-179.
- [138] Sun, Y. X.; Schneider, A.; Jung, Y.; Wang, J.; Dai, J.; Wang, J.; Cook, K.; Osman, N. I.; Koh-Paige, A. J.; Shim, H.; Pienta, K. J.; Keller, E. T.; McCauley, L. K.; Taichman, R. S. Skeletal localization and neutralization of the SDF-1(CXCL12)/CXCR4 axis blocks prostate cancer metastasis and growth in osseous sites *in vivo*. *J. Bone Miner. Res.*, **2005**, *20*(2), 318-329.
- [139] Arvidsson, Y.; Bergström, A.; Arvidsson, L.; Kristiansson, E.; Ahlman, H.; Nilsson, O. Hypoxia stimulates CXCR4 signalling in ileal carcinoids. *Endocr. Relat. Cancer*, **2010**, *17*(2), 303-316.
- [140] Nevo, I.; Sagi-Assif, O.; Meshel, T.; Geminder, H.; Goldberg-Bittman, L.; Ben-Menachem, S.; Shalmon, B.; Goldberg, I.; Ben-Baruch, A.; Witz, I. P. The tumor microenvironment: CXCR4 is associated with distinct protein expression patterns in neuroblastoma cells. *Immunol. Lett.*, **2004**, *92*(1-2), 163-169.
- [141] Burger, M.; Glodek, A.; Hartmann, T.; Schmitt-Graff, A.; Silberstein, L. E.; Fujii, N.; Kipps, T. J.; Burger, J. A. Functional expression of CXCR4 (CD184) on small-cell lung cancer cells mediates migration, integrin activation, and adhesion to stromal cells. *Oncogene*, **2003**, *22*(50), 8093-8101.
- [142] Laird, D. J.; von Andrian, U. H.; Wagers, A. J. Stem cell trafficking in tissue development, growth, and disease. *Cell*, **2008**, *132*(4), 612-630.
- [143] Murakami, T.; Maki, W.; Cardones, A. R.; Fang, H.; Tun Kyi, A.; Nestle, F. O.; Hwang, S. T. Expression of CXC chemokine receptor-4 enhances the pulmonary metastatic potential of murine B16 melanoma cells. *Cancer Res.*, **2002**, *62*(24), 7328-7334.
- [144] Smith, M. C.; Luker, K. E.; Garbow, J. R.; Prior, J. L.; Jackson, E.; Pivnicka-Worms, D.; Luker, G. D. CXCR4 regulates growth of both primary and metastatic breast cancer. *Cancer Res.*, **2004**, *64*(23), 8604-8612.
- [145] Yoon, Y.; Liang, Z.; Zhang, X.; Choe, M.; Zhu, A.; Cho, H. T.; Shin, D. M.; Goodman, M. M.; Chen, Z. G.; Shim, H. CXC chemokine receptor-4 antagonist blocks both growth of primary tumor and metastasis of head and neck cancer in xenograft mouse models. *Cancer Res.*, **2007**, *67*(15), 7518-7524.
- [146] Kajiyama, H.; Shibata, K.; Terauchi, M.; Ino, K.; Nawa, A.; Kikkawa, F. Involvement of SDF-1alpha/CXCR4 axis in the enhanced peritoneal metastasis of epithelial ovarian carcinoma. *Int. J. Cancer*, **2008**, *122*(1), 91-99.
- [147] Kim, S. Y.; Lee, C. H.; Midura, B. V.; Yeung, C.; Mendoza, A.; Hong, S. H.; Ren, L.; Wong, D.; Korz, W.; Merzouk, A.; Salari, H.; Zhang, H.; Hwang, S. T.; Khanna, C.; Helman, L. J. Inhibition of the CXCR4/CXCL12 chemokine pathway reduces the development of murine pulmonary metastases. *Clin. Exp. Metastasis*, **2008**, *25*(3), 201-211.
- [148] Menu, E.; Asosingh, K.; Indraccolo, S.; De Raeye, H.; Van Riet, I.; Van Valckenborgh, E.; Vande Broek, I.; Fujii, N.; Tamamura, H.; Van Camp, B.; Vanderkerken, K. The involvement of stromal derived factor 1alpha in homing and progression of multiple myeloma in the 5TMM model. *Haematologica*, **2006**, *91*(5), 605-612.
- [149] Larsson, O.; Girmata, A.; Girmata, L. Role of insulin-like growth factor 1 receptor signalling in cancer. *Br. J. Cancer*, **2005**, *92*(12), 2097-2101.
- [150] Vasilcanu, R.; Vasilcanu, D.; Rosengren, L.; Natalishvili, N.; Sehat, B.; Yin, S.; Girmata, A.; Axelson, M.; Girmata, L.; Larsson, O. Picropodophyllin induces downregulation of the insulin-like growth factor 1 receptor: potential mechanistic involvement of Mdm2 and beta-arrestin1. *Oncogene*, **2008**, *27*(11), 1629-1638.
- [151] Vitale, L.; Lenzi, L.; Huntsman, S. A.; Canaider, S.; Frabetti, F.; Casadei, R.; Facchin, F.; Carinci, P.; Zannotti, M.; Coppola, D.; Strippoli, P. Differential expression of alternatively spliced mRNA forms of the insulin-like growth factor 1 receptor in human neuroendocrine tumors. *Oncol. Rep.*, **2006**, *15*(5), 1249-1256.
- [152] von Wichert, G.; Jehle, P. M.; Hoeflich, A.; Koschnick, S.; Dralle, H.; Wolf, E.; Wiedenmann, B.; Boehm, B. O.; Adler, G.; Seufferlein, T. Insulin-like growth factor-I is an autocrine regulator of chromogranin A secretion and growth in human neuroendocrine tumor cells. *Cancer Res.*, **2000**, *60*(16), 4573-81.
- [153] Van Gompel, J. J.; Chen, H. Insulin-like growth factor 1 signaling in human gastrointestinal carcinoid tumor cells. *Surgery*, **2004**, *136*(6), 1297-1302.
- [154] Höpfner, M.; Baradari, V.; Huether, A.; Schofl, C.; Scherubl, H. The insulin-like growth factor receptor 1 is a promising target for novel treatment approaches in neuroendocrine gastrointestinal tumours. *Endocr. Relat. Cancer*, **2006**, *13*(1), 135-149.

- [155] Nilsson, O.; Wängberg, B.; Theodorsson, E.; Skottner, A.; Ahlman, H. Presence of IGF-I in human midgut carcinoid tumours--an autocrine regulator of carcinoid tumour growth? *Int. J. Cancer*, **1992**, *51*(2), 195-203.
- [156] Laurent-Puig, P.; Lievre, A.; Blons, H. Mutations and response to epidermal growth factor receptor inhibitors. *Clin. Cancer Res.*, **2009**, *15*(4), 1133-1139.
- [157] Bergmann, F.; Breinig, M.; Hopfner, M.; Rieker, R. J.; Fischer, L.; Kohler, C.; Esposito, I.; Kleeff, J.; Herpel, E.; Ehemann, V.; Friess, H.; Schirmacher, P.; Kern, M. A. Expression pattern and functional relevance of epidermal growth factor receptor and cyclooxygenase-2: novel chemotherapeutic targets in pancreatic endocrine tumors? *Am. J. Gastroenterol.*, **2009**, *104*(1), 171-181.
- [158] Höpfner, M.; Sutter, A. P.; Gerst, B.; Zeitz, M.; Scherubl, H. A novel approach in the treatment of neuroendocrine gastrointestinal tumours. Targeting the epidermal growth factor receptor by gefitinib (ZD1839). *Br. J. Cancer*, **2003**, *89*(9), 1766-75.
- [159] Kidd, M.; Eick, G. N.; Modlin, I. M.; Pfragner, R.; Champaneria, M. C.; Murren, J. Further delineation of the continuous human neoplastic enterochromaffin cell line, KRJ-1, and the inhibitory effects of lanreotide and rapamycin. *J. Mol. Endocrinol.*, **2007**, *38*(1-2), 181-192.
- [160] Nilsson, O.; Wängberg, B.; Kölby, L.; Schultz, G. S.; Ahlman, H. Expression of transforming growth factor alpha and its receptor in human neuroendocrine tumours. *Int. J. Cancer*, **1995**, *60*(5), 645-651.
- [161] Tian, M.; Schiemann, W. P. The TGF-beta paradox in human cancer: an update. *Future Oncol.*, **2009**, *5*(2), 259-271.
- [162] Moustakas, A.; Heldin, C. H. The regulation of TGFbeta signal transduction. *Development*, **2009**, *136*(22), 3699-3714.
- [163] Ishizuka, J.; Beauchamp, R. D.; Sato, K.; Townsend, C. M., Jr.; Thompson, J. C. Novel action of transforming growth factor beta 1 in functioning human pancreatic carcinoid cells. *J. Cell Physiol.*, **1993**, *156*(1), 112-118.
- [164] Wimmel, A.; Wiedenmann, B.; Rosewicz, S. Autocrine growth inhibition by transforming growth factor beta-1 (TGFbeta-1) in human neuroendocrine tumour cells. *Gut*, **2003**, *52*(9), 1308-1316.
- [165] Leu, F. P.; Nandi, M.; Niu, C. The effect of transforming growth factor beta on human neuroendocrine tumor BON cell proliferation and differentiation is mediated through somatostatin signaling. *Mol. Cancer Res.*, **2008**, *6*(6), 1029-1042.
- [166] Kidd, M.; Modlin, I. M.; Pfragner, R.; Eick, G. N.; Champaneria, M. C.; Chan, A. K.; Camp, R. L.; Mane, S. M. Small bowel carcinoid (enterochromaffin cell) neoplasia exhibits transforming growth factor-beta1-mediated regulatory abnormalities including up-regulation of C-Myc and MTA1. *Cancer*, **2007**, *109*(12), 2420-2431.
- [167] Massague, J. TGFbeta in cancer. *Cell*, **2008**, *134*(2), 215-230.
- [168] Bold, R. J.; Ishizuka, J.; Rajaraman, S.; Perez-Polo, J. R.; Townsend, C. M., Jr.; Thompson, J. C. Nerve growth factor as a mitogen for a pancreatic carcinoid cell line. *J. Neurochem.*, **1995**, *64*(6), 2622-2628.
- [169] Kunnimalaiyaan, M.; Ndiaye, M.; Chen, H. Apoptosis-mediated medullary thyroid cancer growth suppression by the PI3K inhibitor LY294002. *Surgery*, **2006**, *140*(6), 1009-1014; discussion 1014-5.
- [170] Nicholson, K. M.; Anderson, N. G. The protein kinase B/Akt signalling pathway in human malignancy. *Cell Signal*, **2002**, *14*(5), 381-395.
- [171] Pitt, S. C.; Chen, H.; Kunnimalaiyaan, M. Phosphatidylinositol 3-kinase-Akt signaling in pulmonary carcinoid cells. *J. Am. Coll. Surg.*, **2009**, *209*(1), 82-88.
- [172] Liu, H.; Radisky, D. C.; Nelson, C. M.; Zhang, H.; Fata, J. E.; Roth, R. A.; Bissell, M. J. Mechanism of Akt1 inhibition of breast cancer cell invasion reveals a protumorigenic role for TSC2. *Proc. Natl. Acad. Sci. USA*, **2006**, *103*(11), 4134-4139.
- [173] Wu, C.; Huang, J. Phosphatidylinositol 3-kinase-AKT-mammalian target of rapamycin pathway is essential for neuroendocrine differentiation of prostate cancer. *J. Biol. Chem.*, **2007**, *282*(6), 3571-83.
- [174] Toulany, M.; Kehlbach, R.; Florczak, U.; Sak, A.; Wang, S.; Chen, J.; Lobrich, M.; Rodemann, H. P. Targeting of AKT1 enhances radiation toxicity of human tumor cells by inhibiting DNA-PKcs-dependent DNA double-strand break repair. *Mol. Cancer Ther.*, **2008**, *7*(7), 1772-1781.
- [175] Davies, H.; Bignell, G. R.; Cox, C.; Stephens, P.; Edkins, S.; Clegg, S.; Teague, J.; Woffendin, H.; Garnett, M. J.; Bottomley, W.; Davis, N.; Dicks, E.; Ewing, R.; Floyd, Y.; Gray, K.; Hall, S.; Hawes, R.; Hughes, J.; Kosmidou, V.; Menzies, A.; Mould, C.; Parker, A.; Stevens, C.; Watt, S.; Hooper, S.; Wilson, R.; Jayatilake, H.; Gusterson, B. A.; Cooper, C.; Shipley, J.; Hargrave, D.; Pritchard-Jones, K.; Maitland, N.; Chenevix-Trench, G.; Riggins, G. J.; Bigner, D. D.; Palmieri, G.; Cossu, A.; Flanagan, A.; Nicholson, A.; Ho, J. W.; Leung, S. Y.; Yuen, S. T.; Weber, B. L.; Seigler, H. F.; Darrow, T. L.; Paterson, H.; Marais, R.; Marshall, C. J.; Wooster, R.; Stratton, M. R.; Futreal, P. A. Mutations of the BRAF gene in human cancer. *Nature*, **2002**, *417*(6892), 949-954.
- [176] Younes, N.; Fulton, N.; Tanaka, R.; Wayne, J.; Straus, F. H., 2nd; Kaplan, E. L. The presence of K-12 ras mutations in duodenal adenocarcinomas and the absence of ras mutations in other small bowel adenocarcinomas and carcinoid tumors. *Cancer*, **1997**, *79*(9), 1804-1808.
- [177] Tannapfel, A.; Vomschloss, S.; Karhoff, D.; Markwarth, A.; Hengge, U. R.; Wittekind, C.; Arnold, R.; Horsch, D. BRAF gene mutations are rare events in gastroenteropancreatic neuroendocrine tumors. *Am. J. Clin. Pathol.*, **2005**, *123*(2), 256-260.
- [178] Karhoff, D.; Sauer, S.; Schrader, J.; Arnold, R.; Fendrich, V.; Bartsch, D. K.; Horsch, D. Rap1/B-Raf signaling is activated in neuroendocrine tumors of the digestive tract and Raf kinase inhibition constitutes a putative therapeutic target. *Neuroendocrinology*, **2007**, *85*(1), 45-53.
- [179] Van Gompel, J. J.; Kunnimalaiyaan, M.; Holen, K.; Chen, H. ZM336372, a Raf-1 activator, suppresses growth and neuroendocrine hormone levels in carcinoid tumor cells. *Mol. Cancer Ther.*, **2005**, *4*(6), 910-917.
- [180] Hall-Jackson, C. A.; Eyers, P. A.; Cohen, P.; Goedert, M.; Boyle, F. T.; Hewitt, N.; Plant, H.; Hedge, P. Paradoxical activation of Raf by a novel Raf inhibitor. *Chem. Biol.*, **1999**, *6*(8), 559-568.
- [181] Chen, H.; Udelsman, R.; Zeiger, M. A.; Ball, D. A. Human achaete-scute homolog-1 is highly expressed in a subset of neuroendocrine tumours. *Oncol. Rep.*, **1997**, *4*, 775-778.
- [182] Lanigan, T. M.; DeRaad, S. K.; Russo, A. F. Requirement of the MASH-1 transcription factor for neuroendocrine differentiation of thyroid C cells. *J. Neurobiol.*, **1998**, *34*(2), 126-134.
- [183] Lo, L. C.; Johnson, J. E.; Wuenschell, C. W.; Saito, T.; Anderson, D. J. Mammalian achaete-scute homolog 1 is transiently expressed by spatially restricted subsets of early neuroepithelial and neural crest cells. *Genes Dev.*, **1991**, *5*(9), 1524-1537.
- [184] Jiang, S. X.; Kameya, T.; Asamura, H.; Umezawa, A.; Sato, Y.; Shinada, J.; Kawakubo, Y.; Igarashi, T.; Nagai, K.; Okayasu, I. hASH1 expression is closely correlated with endocrine phenotype and differentiation extent in pulmonary neuroendocrine tumors. *Mod. Pathol.*, **2004**, *17*(2), 222-229.